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# Shipboard bioassays experiments

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## Objectives

Quantitatively investigate the links between changes in the ocean carbonate system (Ocean Acidification; OA) and:

- natural microbial community physiology and morphometry (including  $\text{CaCO}_3$  shells),
- plankton biodiversity and community structure,
- biogeochemical rates,
- food webs and
- climate-relevant processes.

# Containerised lab facility

## Controlled conditions in a refrigerated container:

Temperature adjusted to match with the *in situ*

Refrigeration unit

Artificial light sources:  $100 \mu\text{E m}^{-2} \cdot \text{s}^{-1}$  (14/8 light/dark cycle)

White light (LED panels)

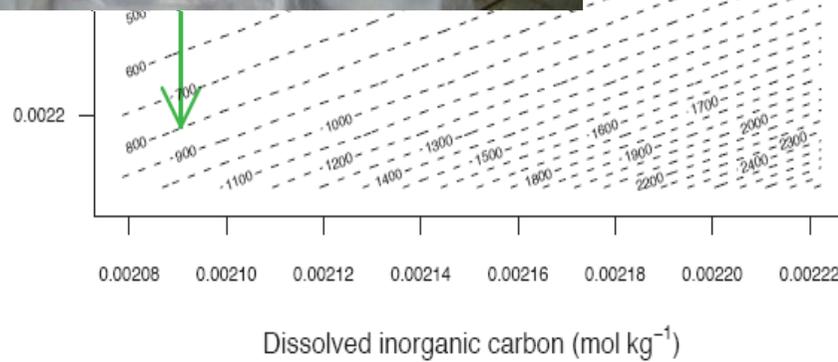


# Set-up

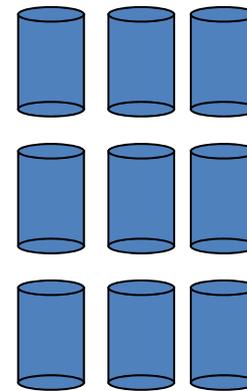


TD of water, 24 x 20L = 480L,  
 er into bottles on deck

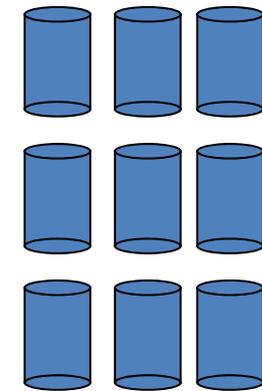
$\text{HCO}_3^- + \text{HCl}$  to achieve target  $\text{pCO}_2$



*m*

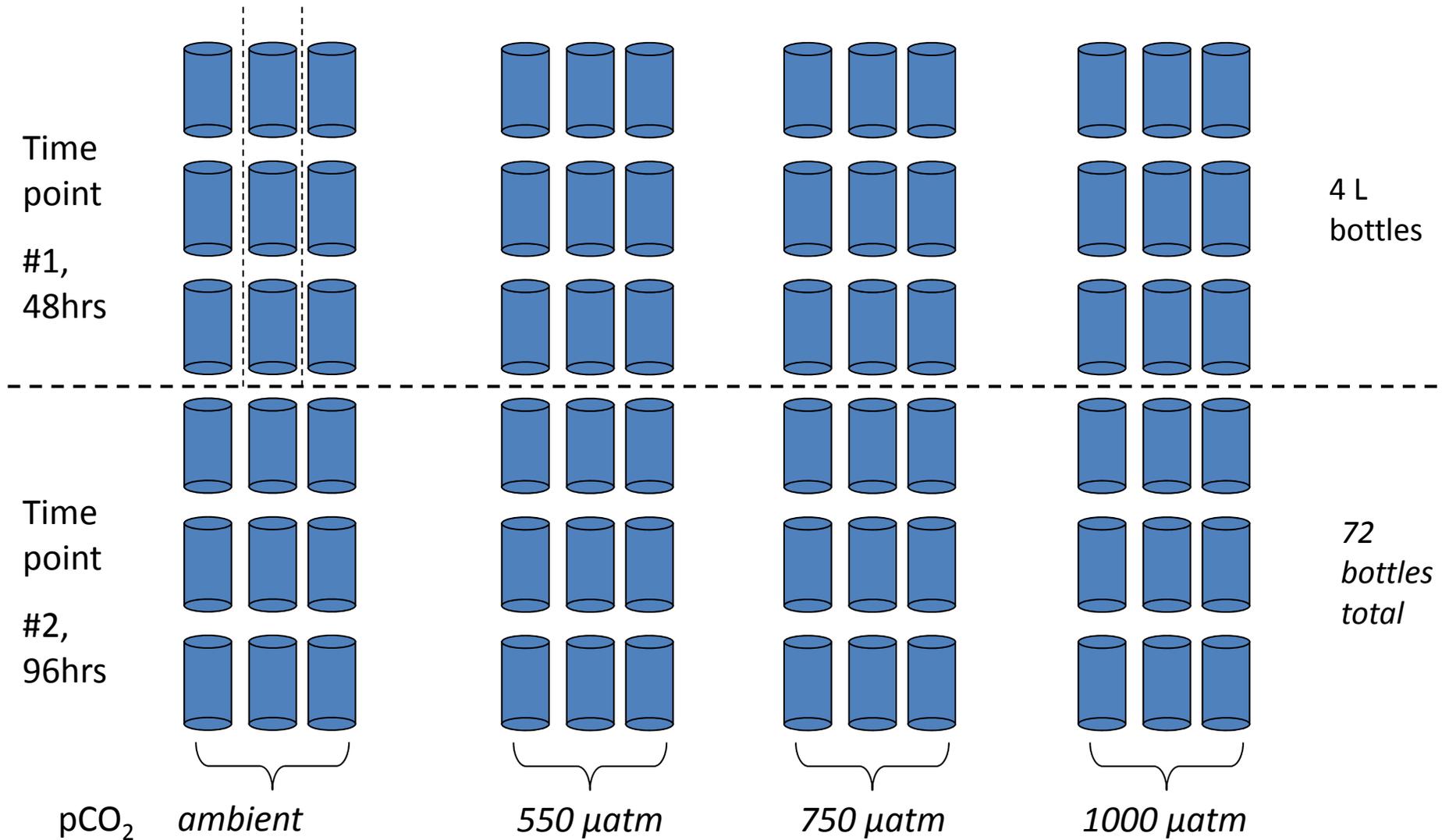


*750 µatm*



*1000 µatm*

# Set-up



# Cruise track (June-July 2011) and bioassay locations

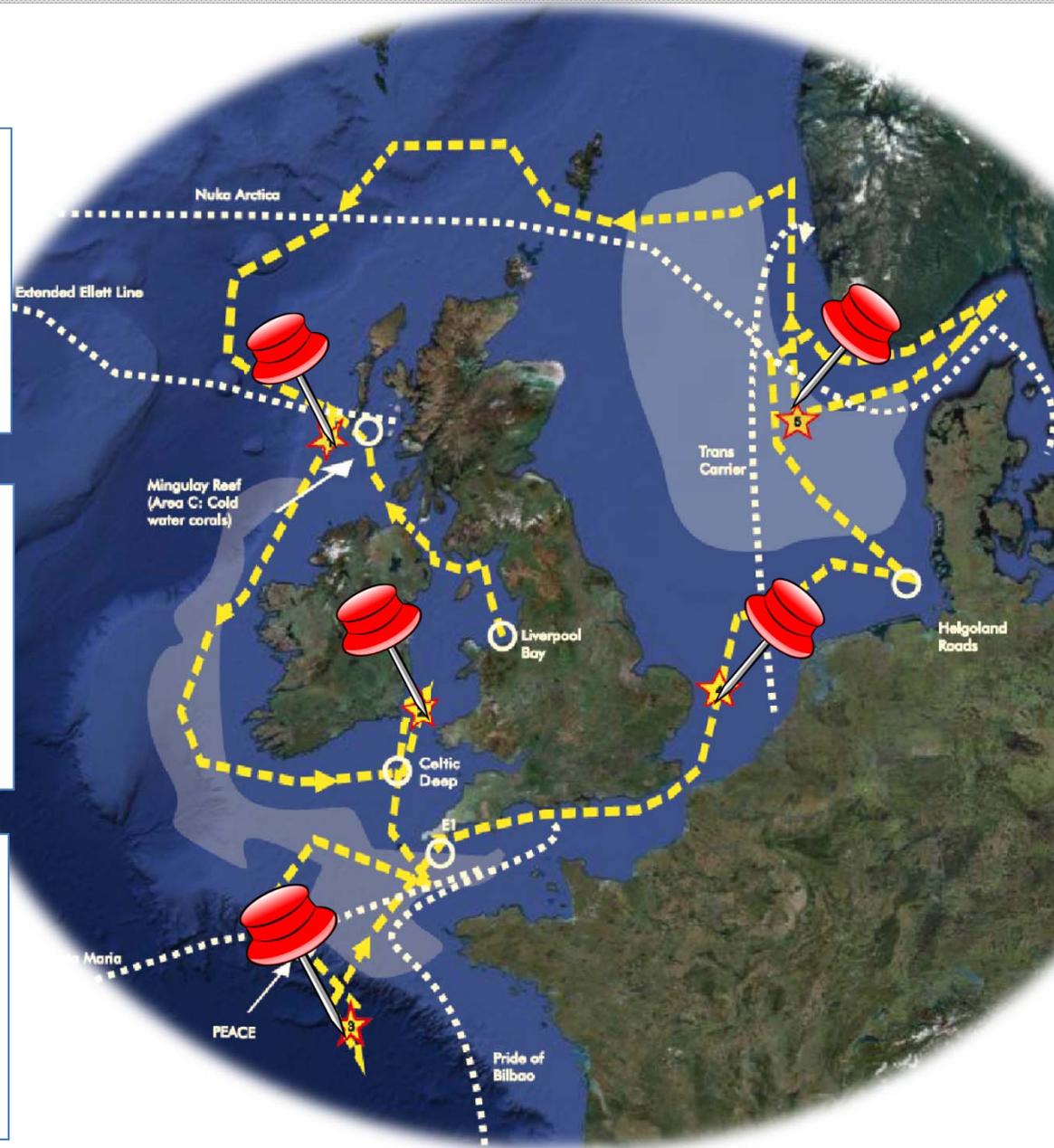
**E01** Stratified  
 T° C: 11.27  
 Salinity: 34.8  
 pCO<sub>2</sub>: 338  
 DIN: 1 μM  
 Si: 2 μM  
 P: 0.09 μM

**E02** Mixed  
 T° C: 11.77  
 Salinity: 34.44  
 pCO<sub>2</sub>: 336  
 DIN: 0.3 μM  
 Si: 0.44 μM  
 P: 0.14 μM

**E03** Stratified  
 T° C: 15.31  
 Salinity: 35.77  
 pCO<sub>2</sub>: 342  
 DIN: 0.56 μM  
 Si: 0.6 μM  
 P: 0.05 μM

**E05** Stratified  
 T° C: 13.96  
 Salinity: 35  
 pCO<sub>2</sub>: 370  
 DIN: 1.7 μM  
 Si: 0.04 μM  
 P: 0.06 μM

**E04** Mixed  
 T° C: 14.57  
 Salinity: 34.05  
 pCO<sub>2</sub>: 407  
 DIN: 0.8 μM  
 Si: 0.8 μM  
 P: 0.13 μM



## Measured parameters

**Carbonate chemistry** (*UoSoton*)

**Macro-nutrients** (*NOCS*)

**POC/PON/POP** (*UoSoton*)

**Total Chl<sub>a</sub>/size fractionated chlorophyll<sub>a</sub>** (*UoSoton, UoEssex*)

**Photophysiology** (*Fv/Fm, σ, τ*) (*UoEssex*)

**P/E** (*UoEssex*)

**Primary production** (*NOCS*)

**Calcite production** (*NOCS*)

**Coccolithophores assemblage** (*NHM*)

**PIC** (*NHM*)

**Microbial community composition** (*NOCS, PML*)

**Respiration** (*UoSoton*)

**Ammonium** (*UoSoton*)

**DOC/TEP** (*UoSoton*)

**DMSP production, DMSP-DMSP conversion rates** (*PML*)

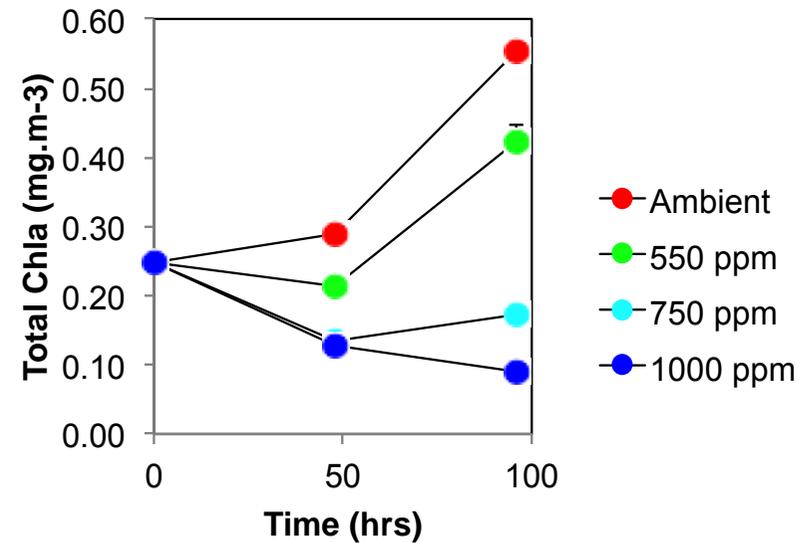
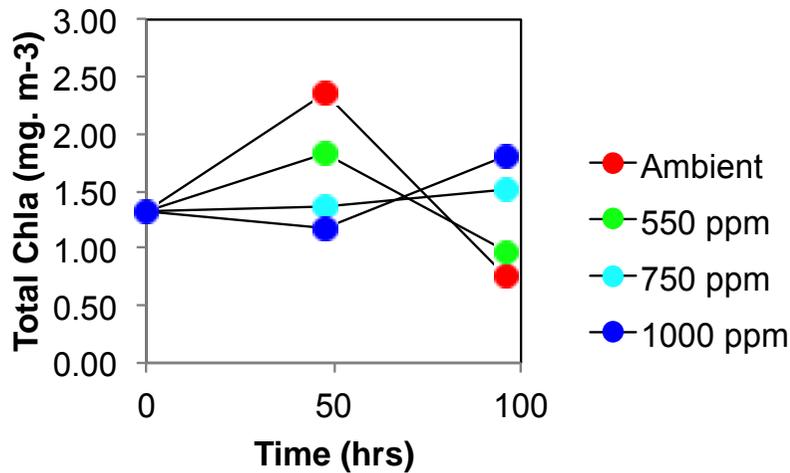
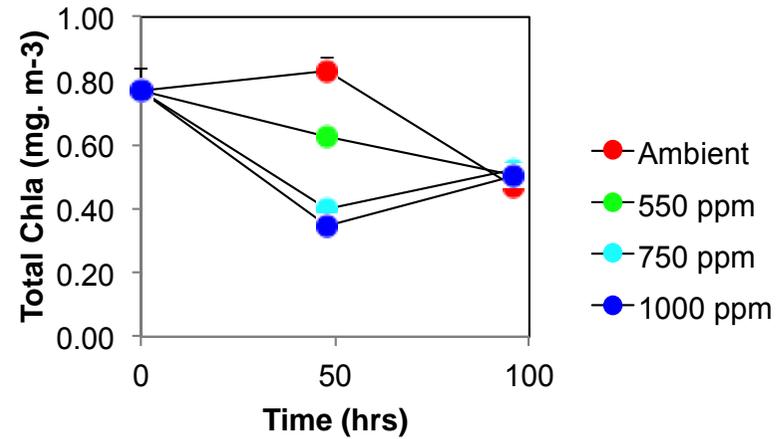
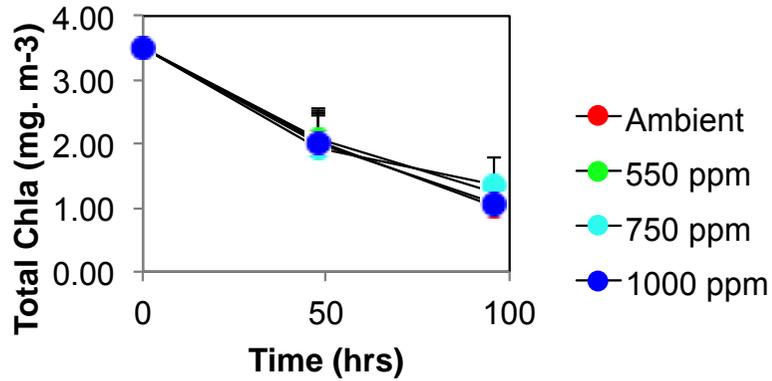
**CH<sub>4</sub>, N<sub>2</sub>O** (*PML*)

**Oxidation rates of NH<sub>4</sub><sup>+</sup> and NO<sub>2</sub><sup>-</sup>** (*PML*)

**Genetic diversity** (*MBA*)

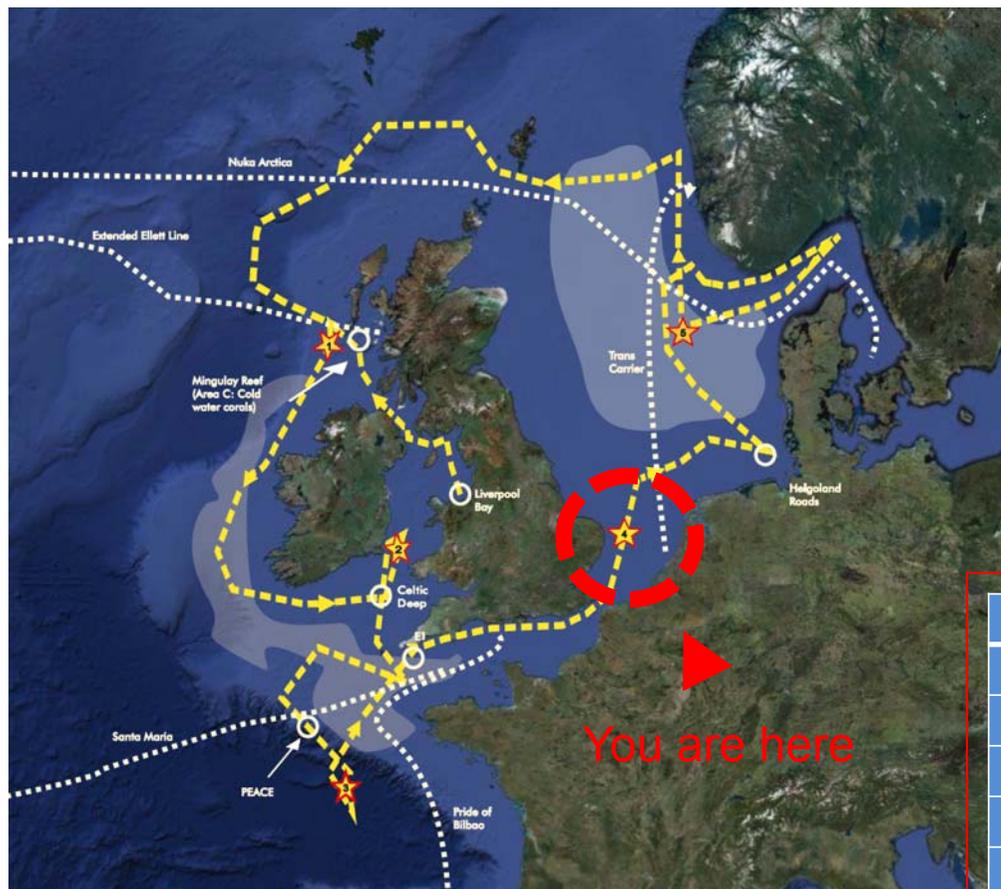
**Proteins/RNA** (*UoSoton*)

# Diversity in responses to ocean acidification



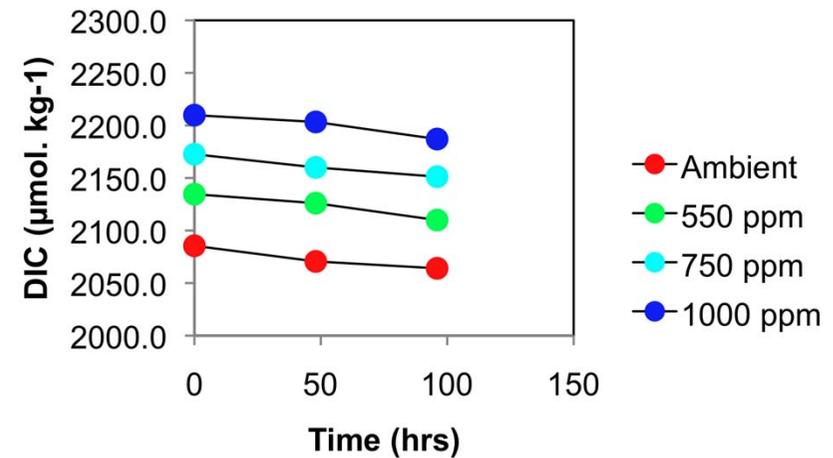
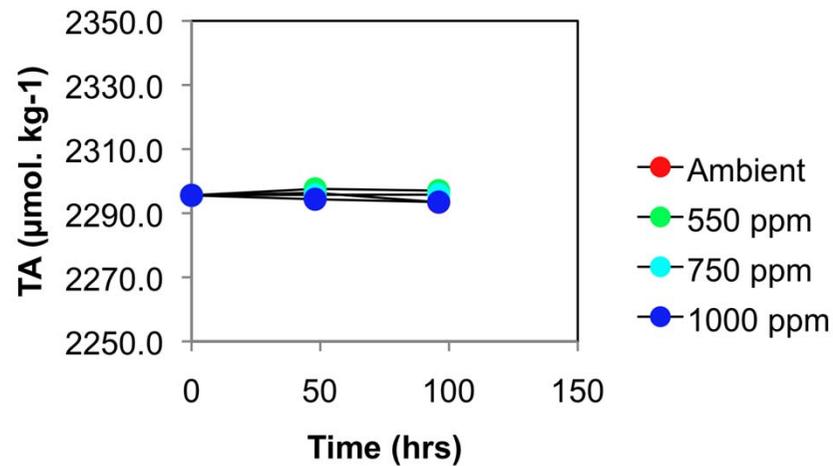
Discrepancies in Chl<sub>a</sub> trends between bioassays illustrate the importance of initial conditions in response to OA.

## Zoom on the South North sea bioassay (E04)



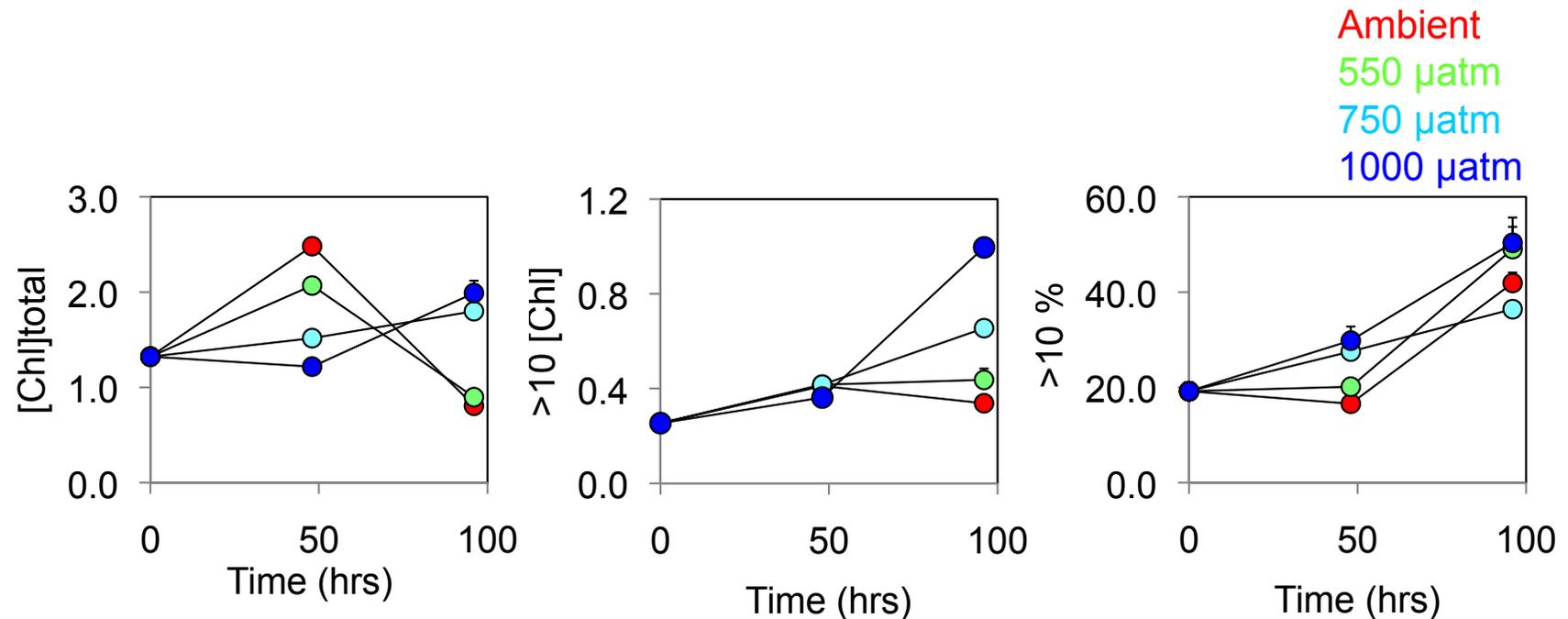
Latitude [deg+veN]	52.59
Longitude [deg+veW]	2.29
Water column	Mixed
Temperature (°C)	14.57
Salinity	34.05
pCO <sub>2</sub> (ppm)	407
Nitrates/nitrites (μM)	0.8
Phosphates (μM)	0.13
Silicates (μM)	0.8

## Carbonate chemistry in E04 bioassay



As expected, the TA remained unchanged with time and pCO<sub>2</sub> through the experiment while the DIC concentration is increasing with pCO<sub>2</sub> and decreasing with time.

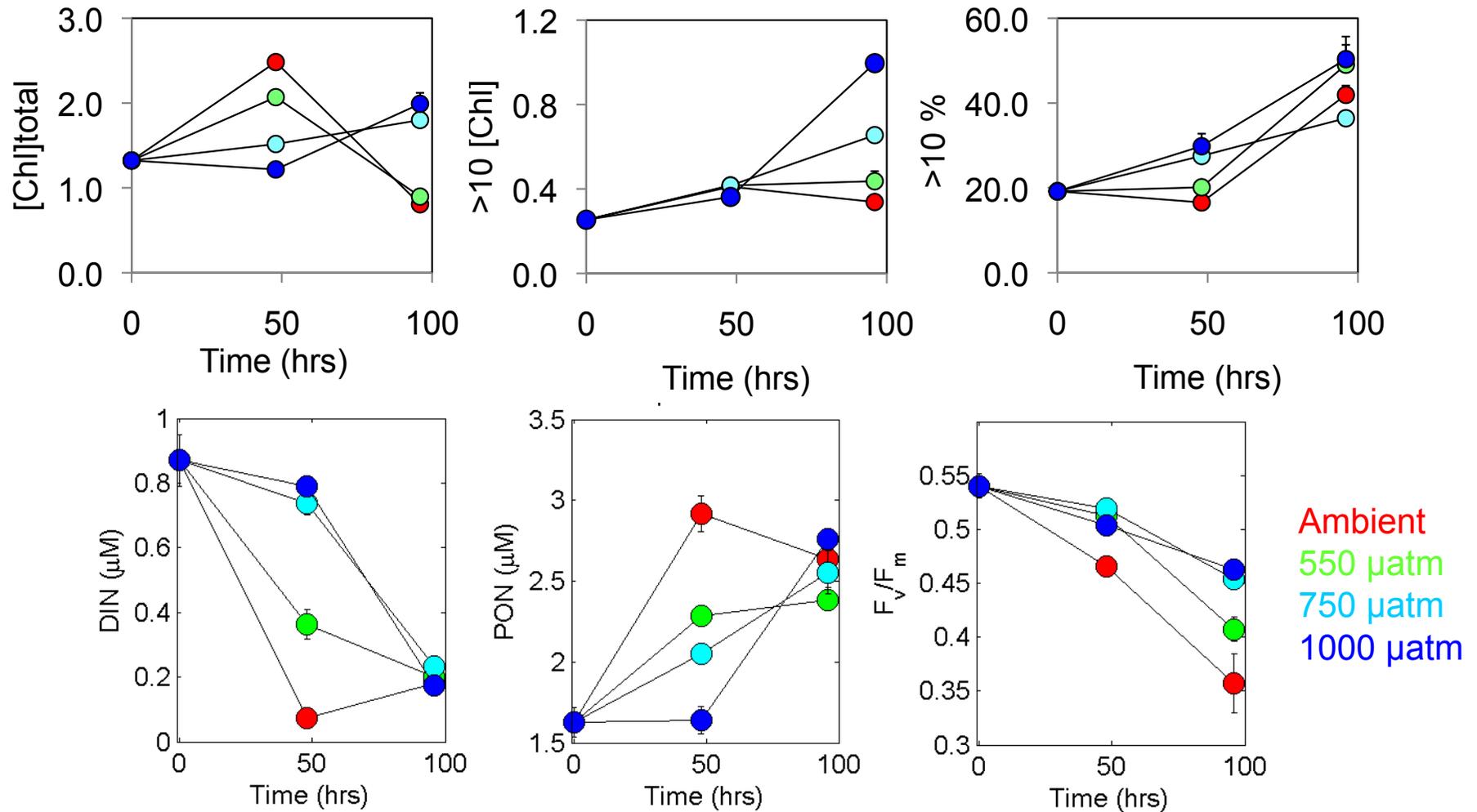
# Interpretation of Time series experiments



Highest Chl<sub>a</sub> values for the ambient condition after 2 days while the bloom is delayed with increasing pCO<sub>2</sub>.

The small size phytoplankton dominates the microbial community (80% at T<sub>0</sub>) and appears more sensitive to ocean acidification than > 10 μm fraction.

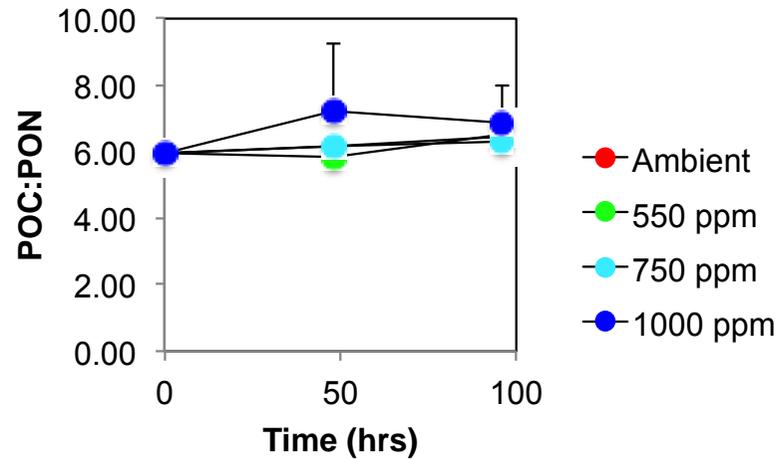
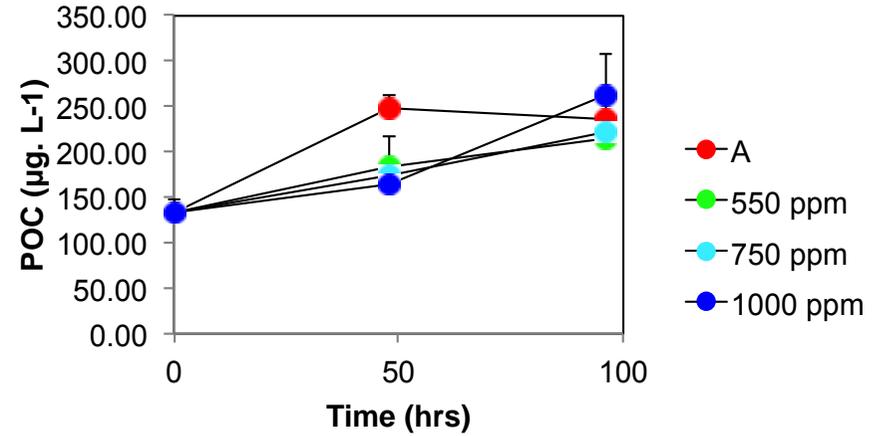
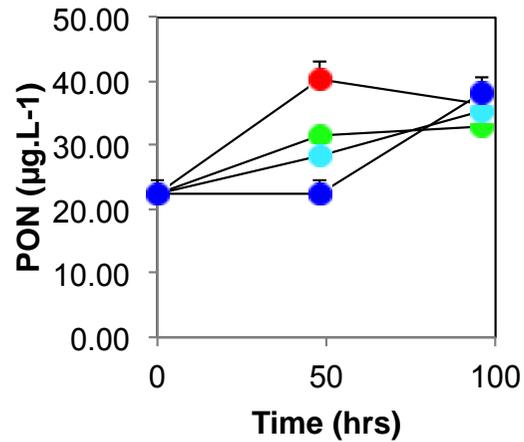
# Interpretation of Time series experiments



Slow down in nutrients uptake and organic matter production with increasing pCO<sub>2</sub>.

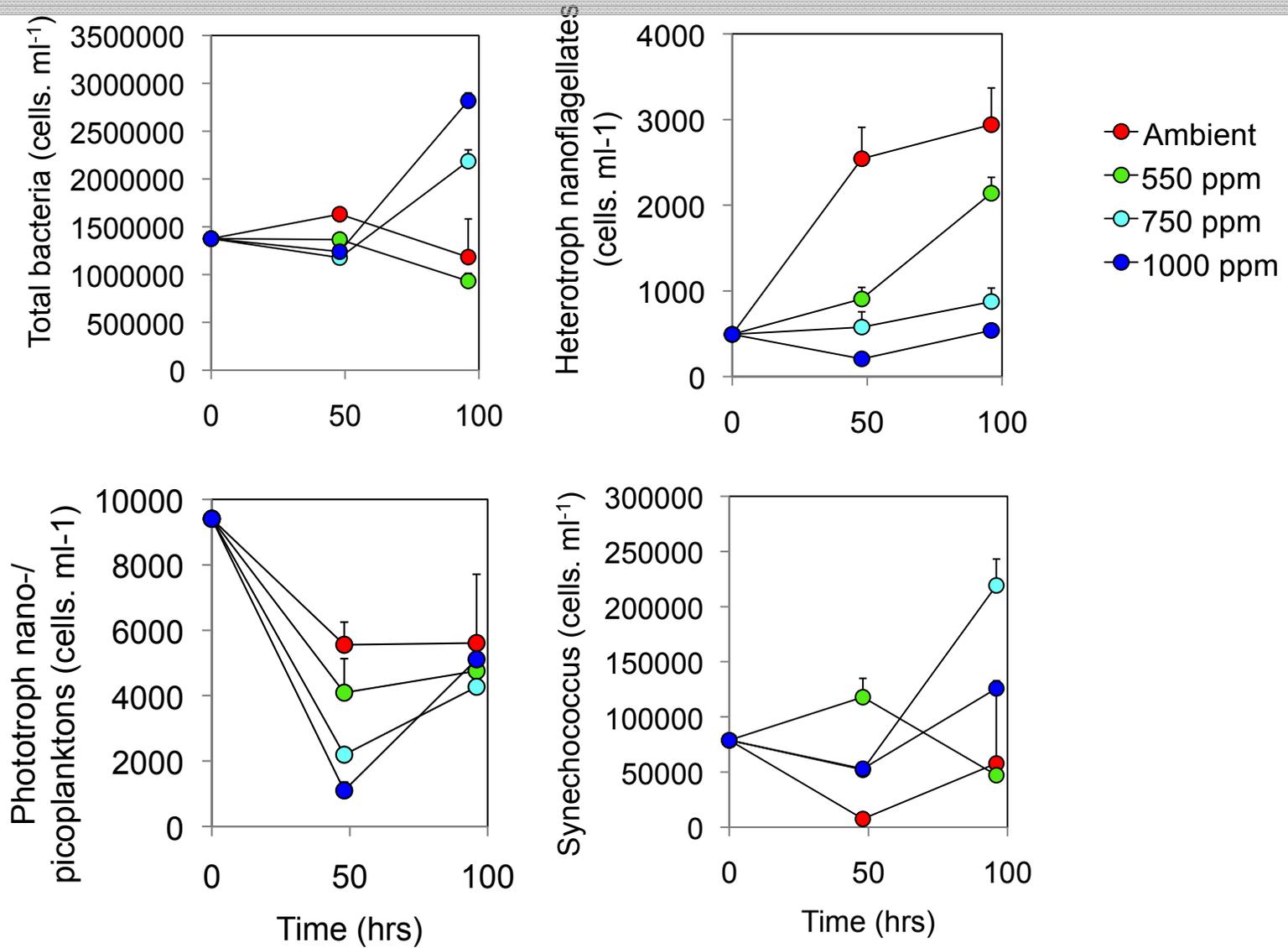
Increase in photo-efficiency with increasing pCO<sub>2</sub> (specific to this bioassay).

# Interpretation of Time series experiments



The production of organic matter is affected by high pCO<sub>2</sub> while C:N ratio remains unchanged.

# Interpretation of Time series experiments



Shift in community composition with increasing pCO<sub>2</sub>.

## Conclusions

The natural communities observed were impacted by ocean acidification.

After 2 days, the chlorophyll content is higher under low  $p\text{CO}_2$ . The peak of chlorophyll corresponds to the highest particulate organic matter concentration and the exhaustion of nutrient sources.

After 4 days, the trend is inverted. As the nutrients uptake is lower at high  $p\text{CO}_2$ , the bloom is delayed and occurs later.

C:N stoichiometry was not affected by higher  $p\text{CO}_2$  and similar to the Redfield ratio of 6.6 in today's ocean.

Shift in microbial community structure (increase in bacteria and decrease in small size fraction and nanoflagellates) with changing  $p\text{CO}_2$ .

## Take home messages

All the measurements performed during the cruise are required to interpret correctly the data from each bioassay,

A strong group-specific response to high  $p\text{CO}_2$  was observed in the majority of bioassay experiments.

Such responses have important implications for trophic dynamics and biogeochemical cycling in both present day and future oceanic ecosystems

The diversity of initial environmental conditions as well as the multidisciplinary approach adopted as part of the project will potentially highlight new acclimation mechanisms of the planktonic community to future ocean acidification.

# Acknowledgments



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