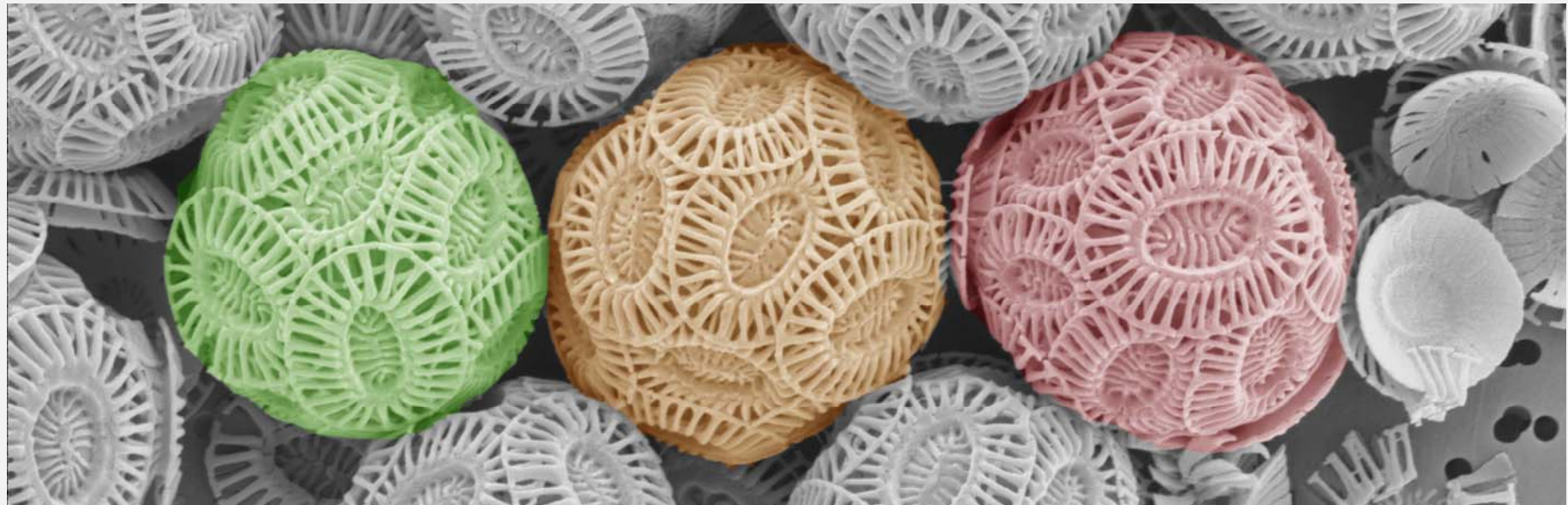
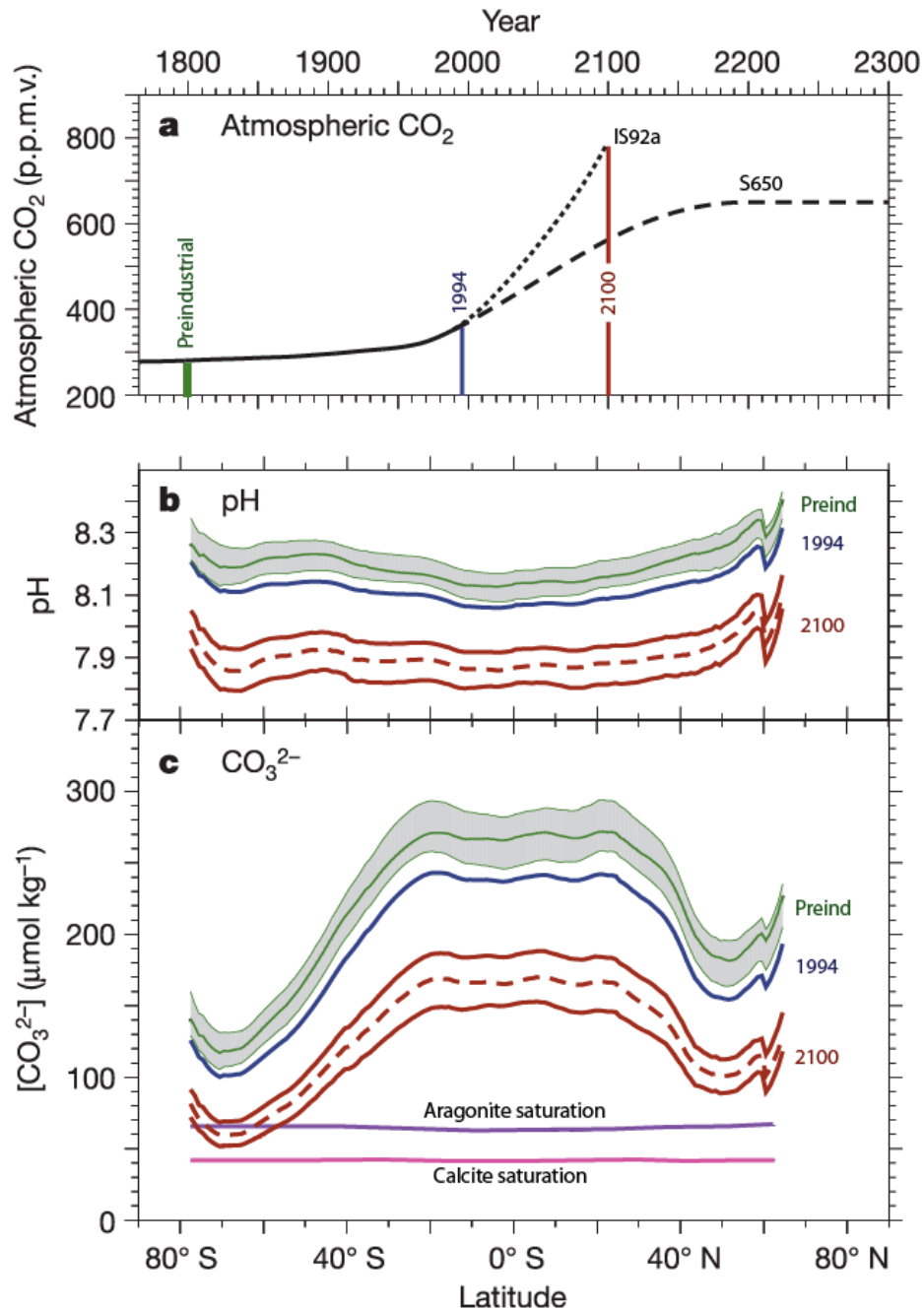


Activities, outputs and outcomes from the UKOA sea surface consortium

## Coccolithophorid distributions and Bioassay Responses

*Jeremy Young, University College London  
Alex Poulton, Toby Tyrell, NOC Southampton  
and many others*

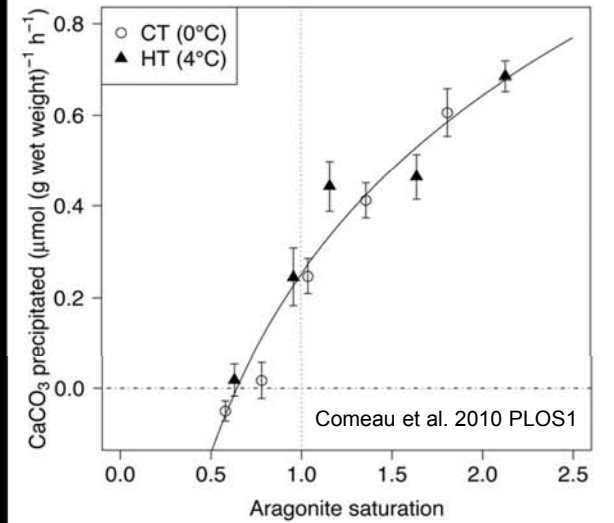
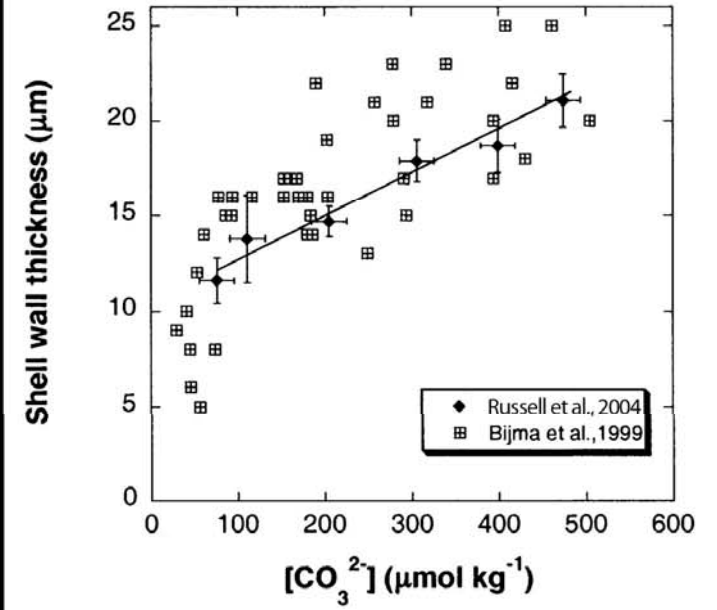
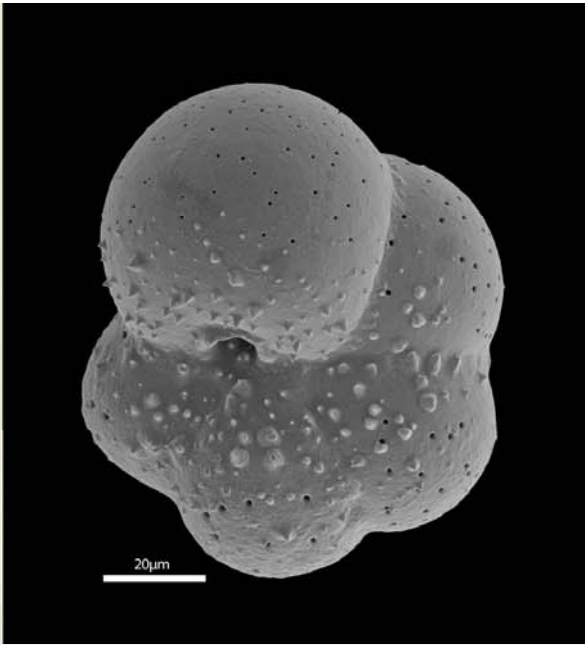
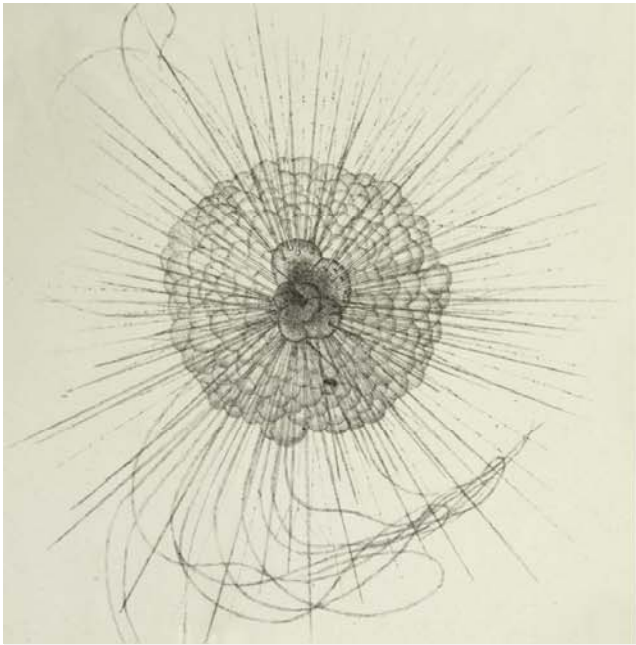




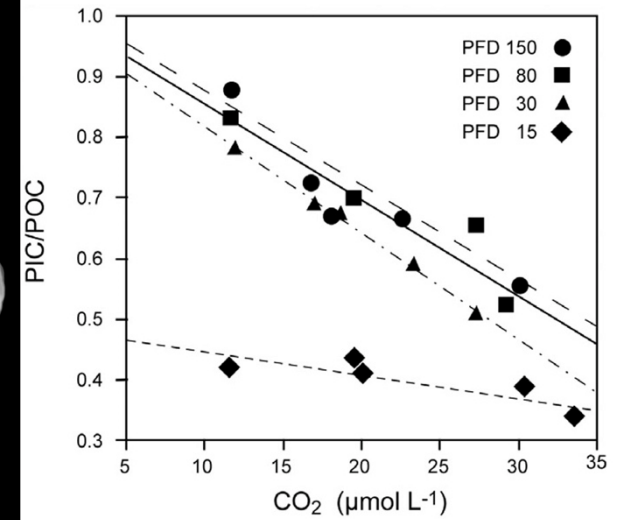
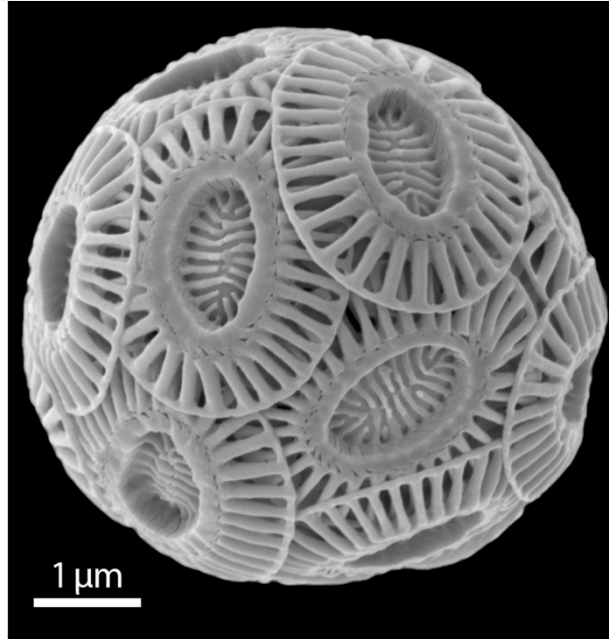
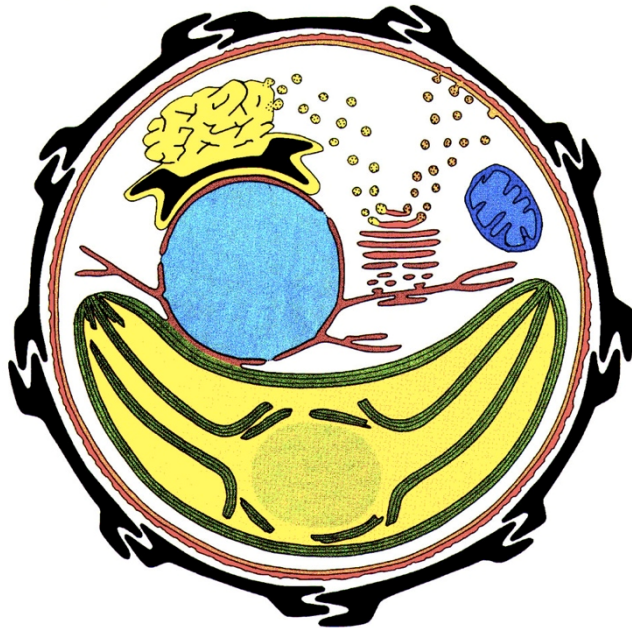
## Hypotheses

*Overarching:* that OA will have significant deleterious impacts on the oceanic ecosystem at CO<sub>2</sub> levels below those which will cause catastrophic global warming

*More specific:* that calcification by coccolithophores is sensitive to change in degree of calcite super-saturation in order of  $\Omega_{\text{calcite}} = 3-5$

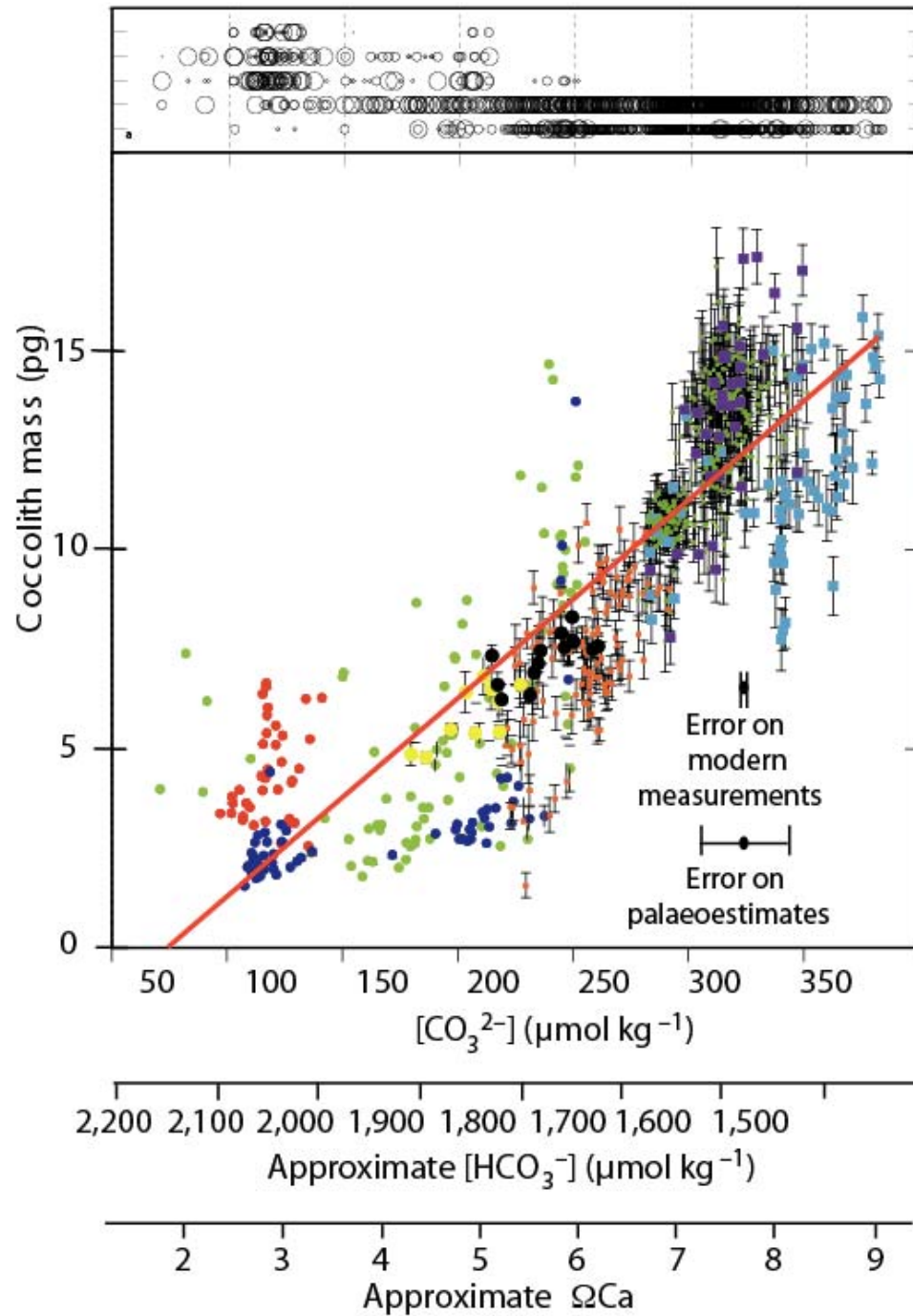


calcification by forams and pteropods does seem to be sensitive to change in degree of super-saturation



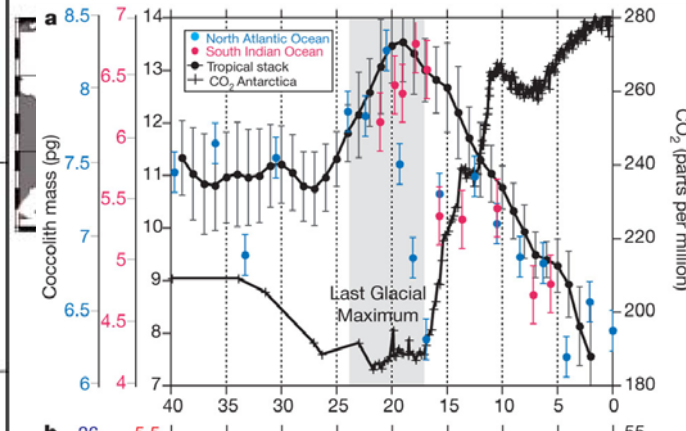
Riebesel et al 2004 J Oceanography

some lab work suggests calcification by coccolithophores is sensitive to change in degree of super-saturation



Malformed  
 E. huxleyi R  
 E. huxleyi C  
 E. huxleyi A  
 G. oceanica

Beaufort *et al.* 2011 *Nature*



some geological and plankton data suggests strong impact of calcite saturation on coccolithophore calcification and ecology

**Primary Effects (Hypothesized)**

**Secondary Effects**

**Tertiary Effects**

CaCO<sub>3</sub> Shells

Inhibition of calcification/  
calcifying plankton

Biogeochemical Rates

Stimulus of photosynthesis &  
varied effects on other  
biogeochemical processes

changes in plankton  
community structure

Community

Biogeochemistry

changes in oceanic  
biogeo-chemistry

Climate

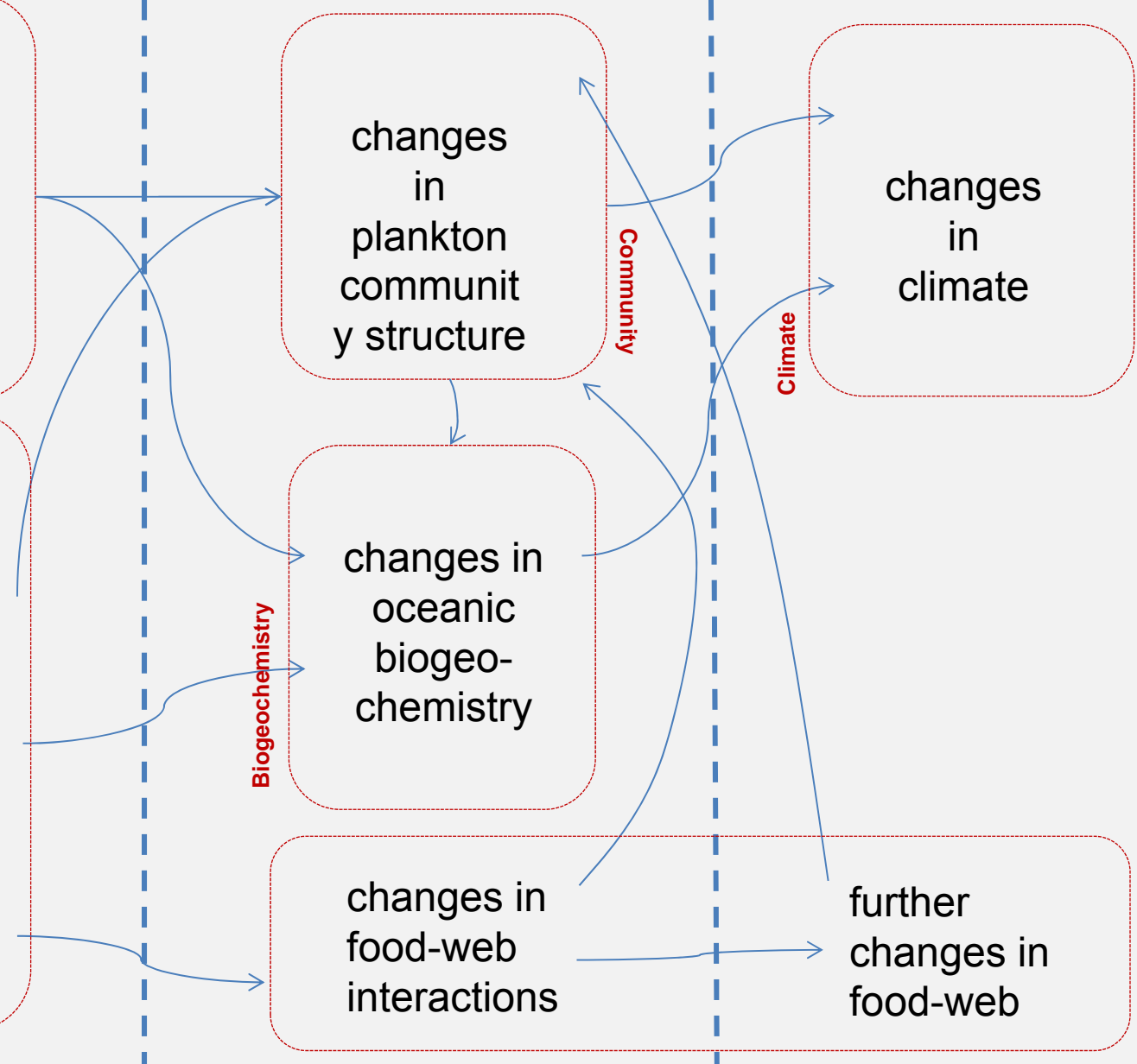
changes in  
climate

changes in  
food-web  
interactions

Food Web

further  
changes in  
food-web

How will ocean acidification affect the plankton ecosystem and the services it provides?



## Primary Effects (Hypothesized)

## Secondary Effects

## Tertiary Effects

CaCO<sub>3</sub> Shells

- H: OA → coccolithophore morphology  
T: examine coccospheres under SEM, morphometrics
- H: OA → pteropod morphology  
T: examine pteropod shells under SEM, measure SNW
- H: OA → foraminifera morphology  
T: examine foram shells under SEM, measure SNW

Biogeochemical Rates

- H: OA → increased photosynthetic rate (some groups)  
T: measure rate of <sup>14</sup>C uptake into POC
- H: OA → increased respiration rate  
T: measure O<sub>2</sub> accumulation in dark bottles
- H: OA → nitrification rate  
T: measure <sup>15</sup>NH<sub>4</sub> uptake
- H: OA → reduced calcification rate  
T: measure rate of <sup>14</sup>C uptake into PIC
- H: OA → increased DOC production, elevated C:N & C:P  
T: measure DOC, POC, PON, POP
- H: OA → altered DMSP cycling  
T: measure in-vivo DMSP synthesis, DMSP consumption

Biogeochemistry

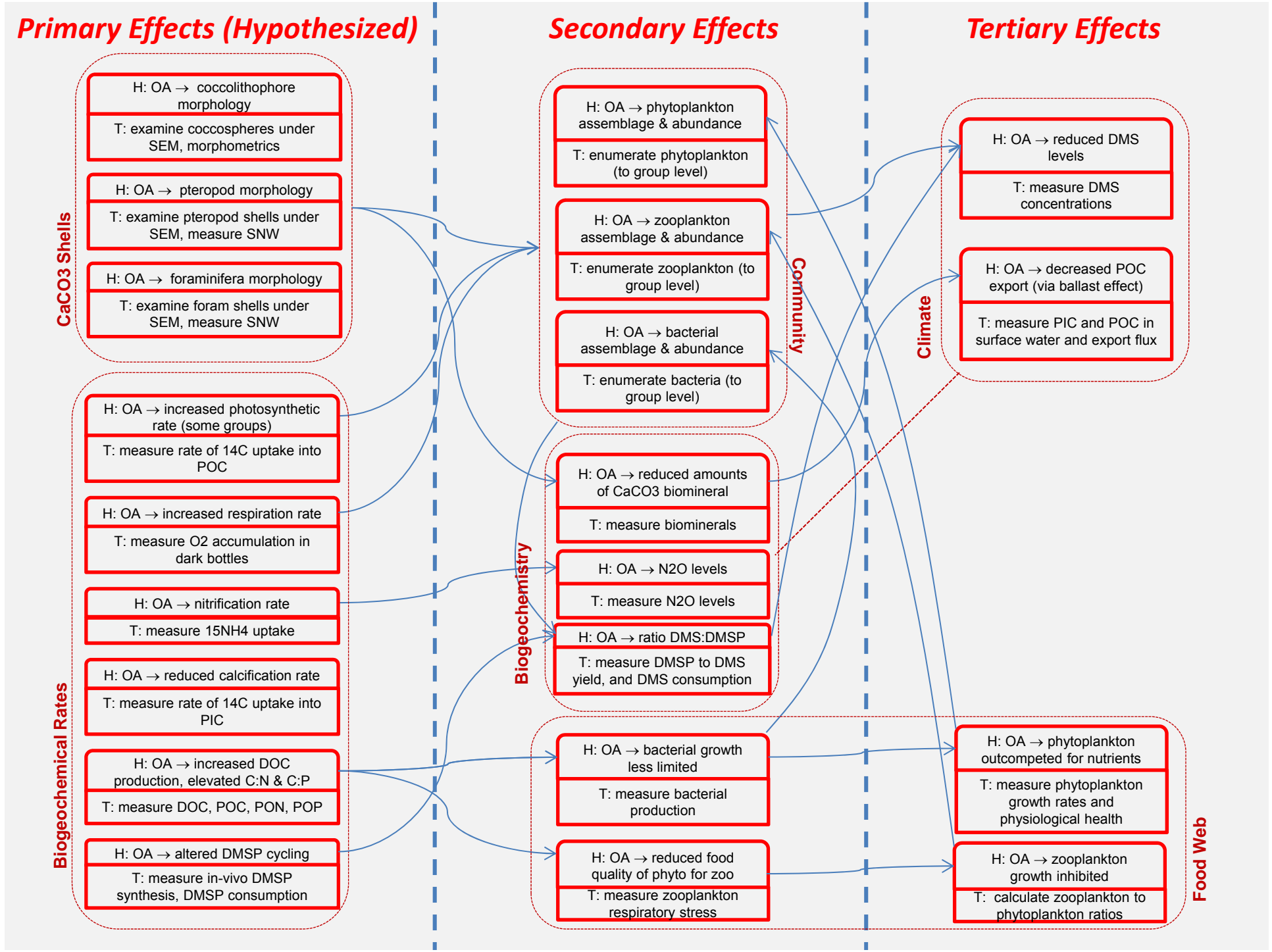
- H: OA → phytoplankton assemblage & abundance  
T: enumerate phytoplankton (to group level)
- H: OA → zooplankton assemblage & abundance  
T: enumerate zooplankton (to group level)
- H: OA → bacterial assemblage & abundance  
T: enumerate bacteria (to group level)
- H: OA → reduced amounts of CaCO<sub>3</sub> biomineral  
T: measure biominerals
- H: OA → N<sub>2</sub>O levels  
T: measure N<sub>2</sub>O levels
- H: OA → ratio DMS:DMSP  
T: measure DMSP to DMS yield, and DMS consumption
- H: OA → bacterial growth less limited  
T: measure bacterial production
- H: OA → reduced food quality of phyto for zoo  
T: measure zooplankton respiratory stress

Community

Climate

Food Web

- H: OA → reduced DMS levels  
T: measure DMS concentrations
- H: OA → decreased POC export (via ballast effect)  
T: measure PIC and POC in surface water and export flux
- H: OA → phytoplankton outcompeted for nutrients  
T: measure phytoplankton growth rates and physiological health
- H: OA → zooplankton growth inhibited  
T: calculate zooplankton to phytoplankton ratios





Cruise D366  
June-July 2011

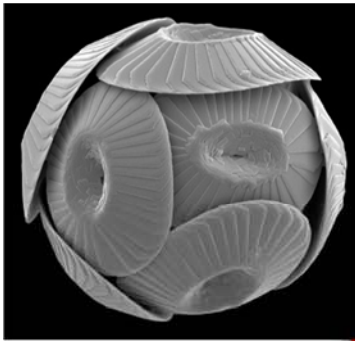
5 Bioassay  
experiments

72 CTD stations

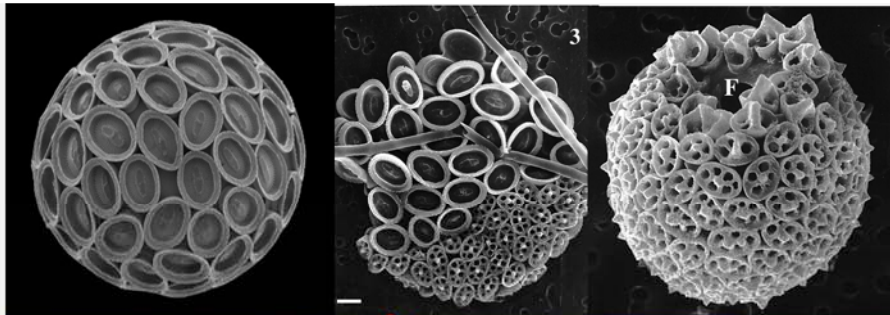
320 Underway  
samples



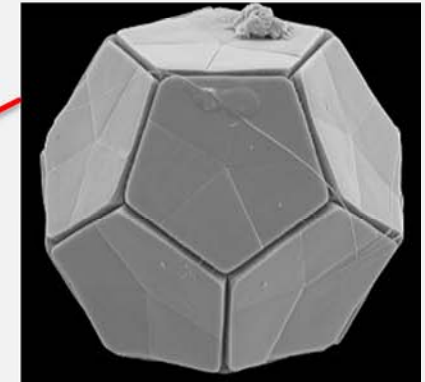




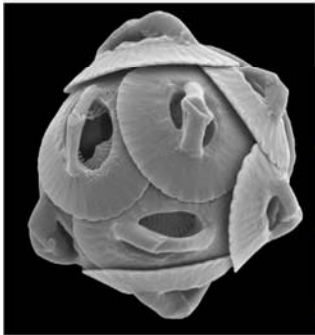
*Coccolithus pelagicus*



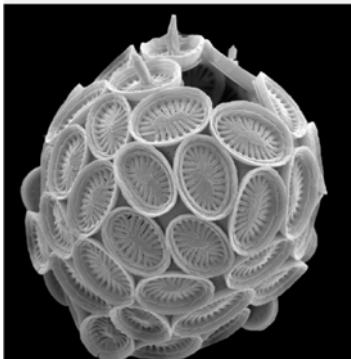
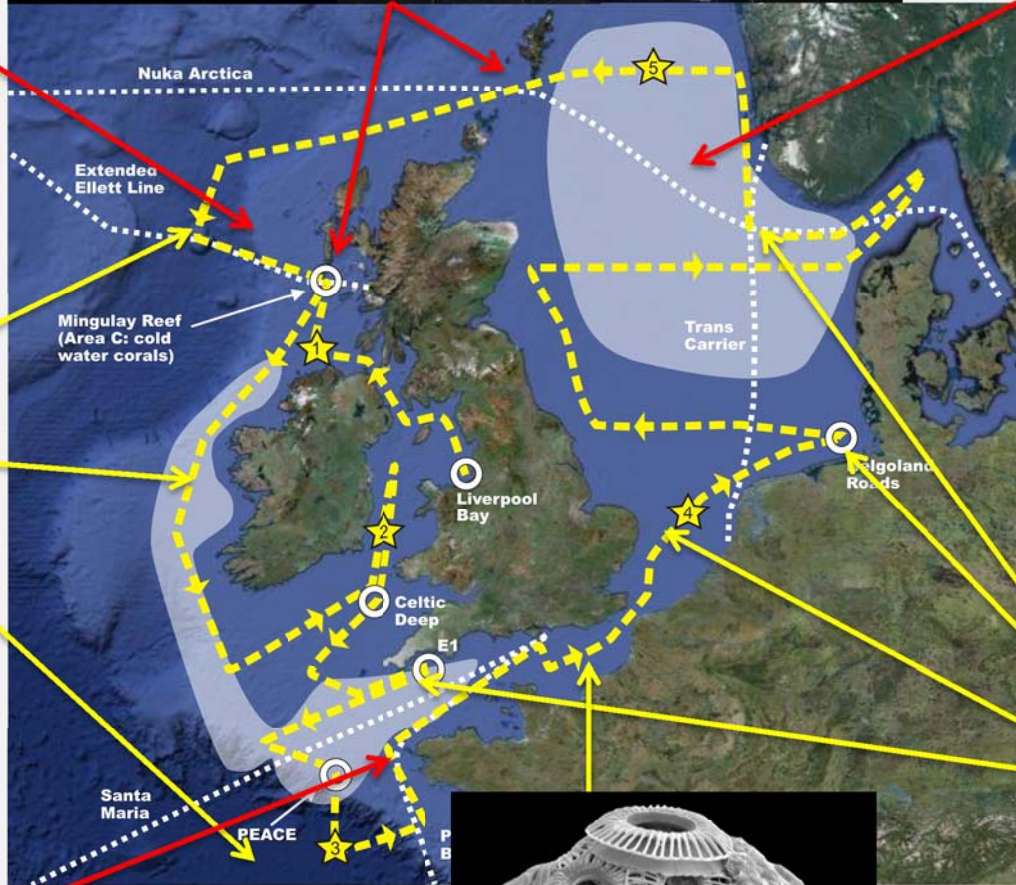
*Coronosphaera mediterranea*



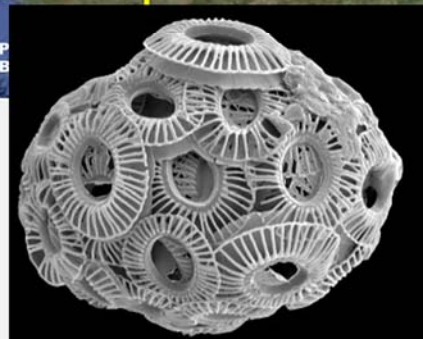
*Braarudosphaera bigelowii*



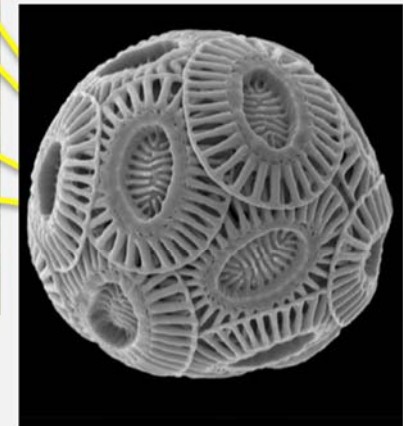
*Gephyrocapsa muelleriae*



*Syracosphaera* sp.



*Emiliana huxleyi* type B



*Emiliana huxleyi* type A

# Cruise JR271

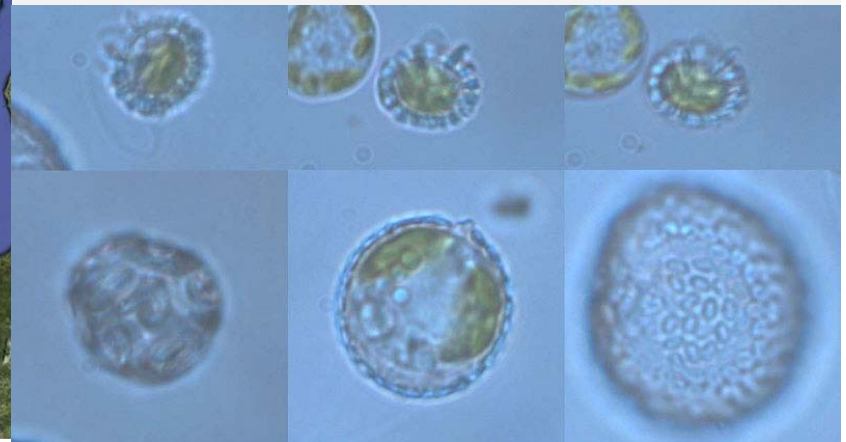
June-July 2012

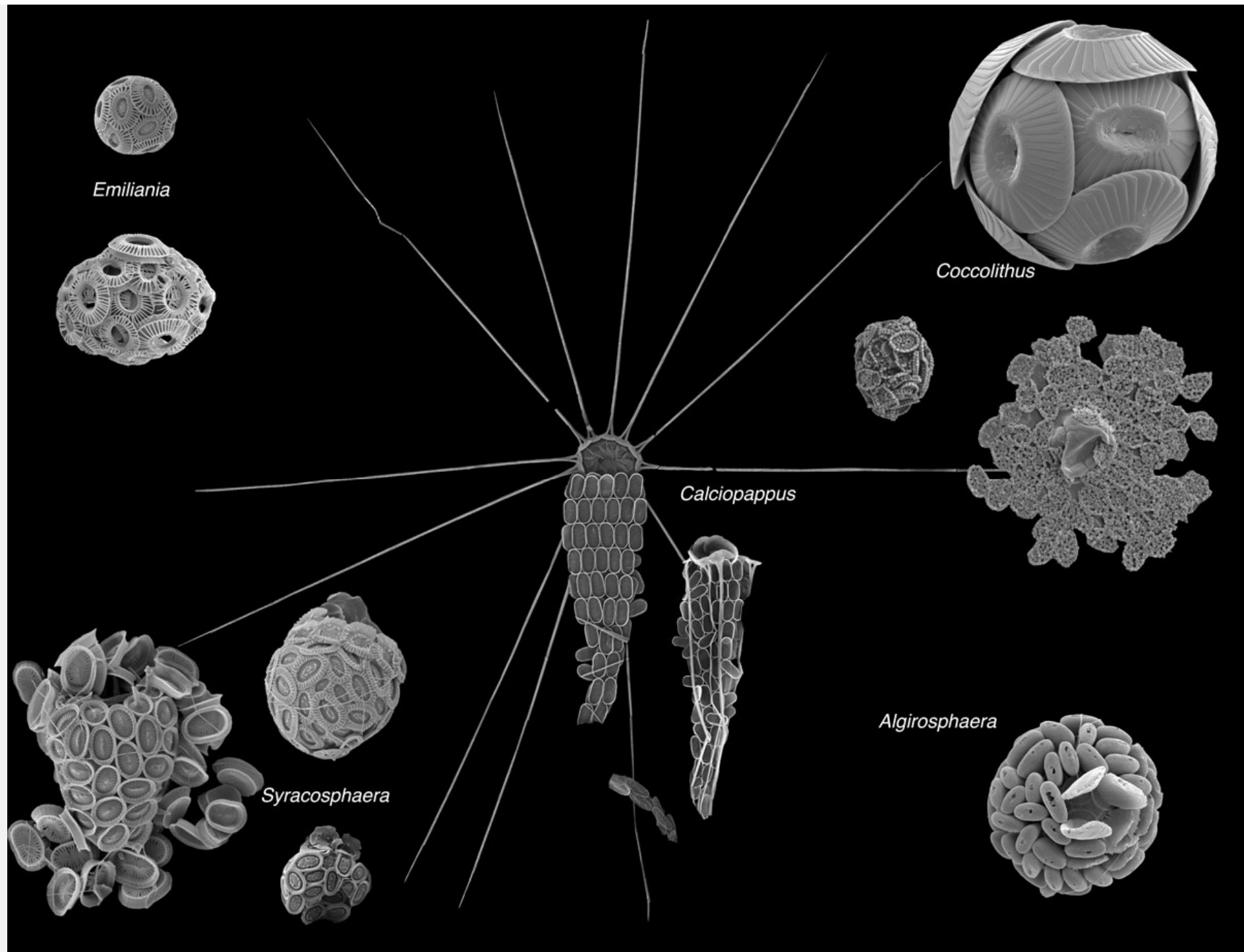
40 CTD stations (usually 6 depths)

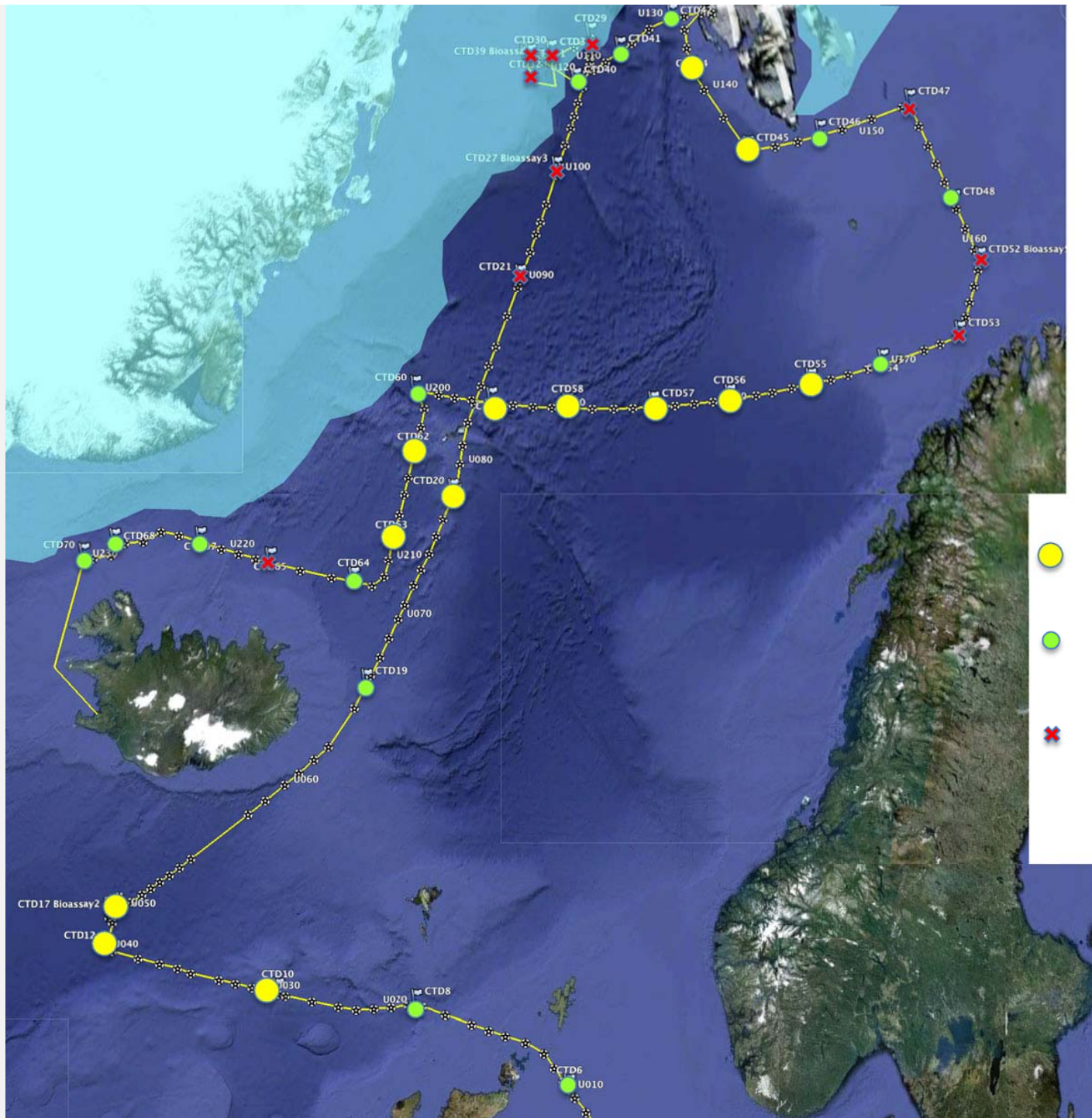
230 underway samples

5 bioassays (but coccos only in 1 & 2)

lots of ice

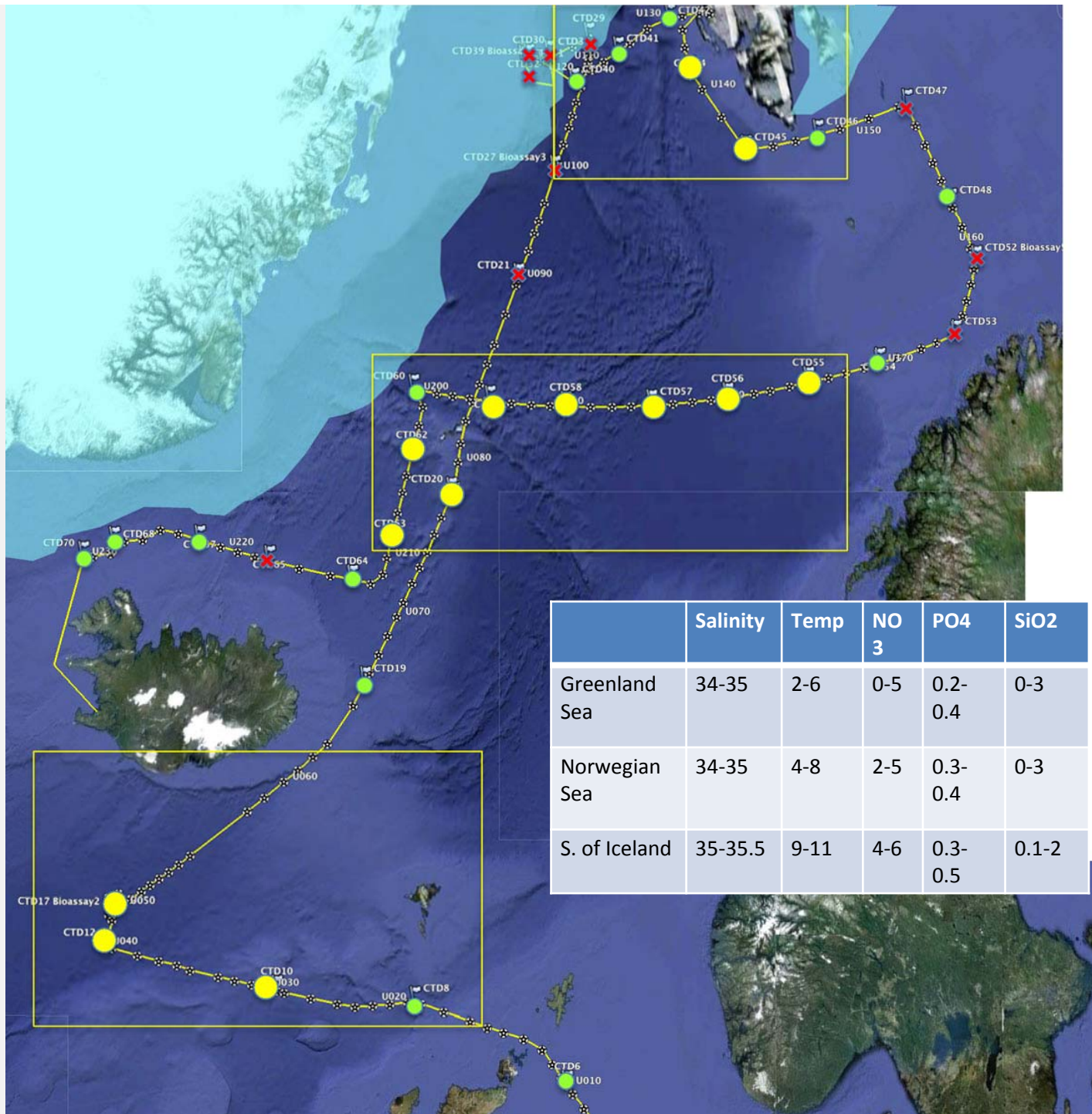






## JR 271 coccolithophore abundance

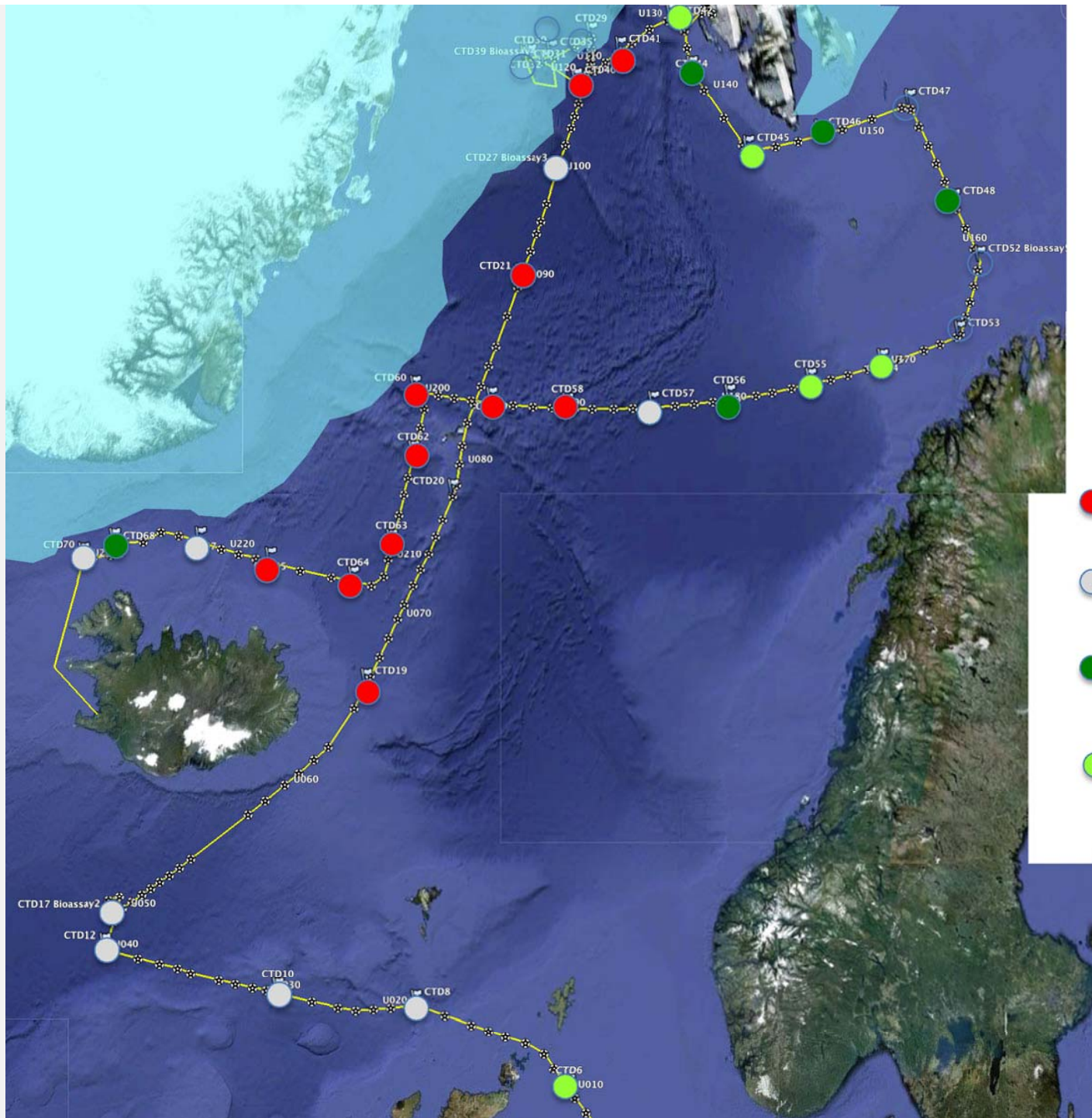
- >100,000 cells/l
- 10-100,000 cells/l
- × <10,000 cells/l



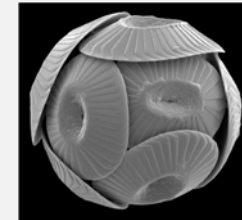
Greenland Sea  
*Emiliana*  
***Coccolithus* HOL**  
*Algirosphaera*

Norwegian Sea  
*Emiliana*  
*Calciopappus*  
*Coccolithus* HET +HOL  
*Algirosphaera*

South of Iceland  
*Emiliana*  
*Calciopappus*  
*Coccolithus* HET  
***Syracosphaera***



JR 271  
*Emiliana:Coccolithus*



- Ehux < 5x Cpel
- Ehux > 5x Cpel
- Ehux > 50x Cpel
- Ehux > 500x Cpel
- sample +/- barren

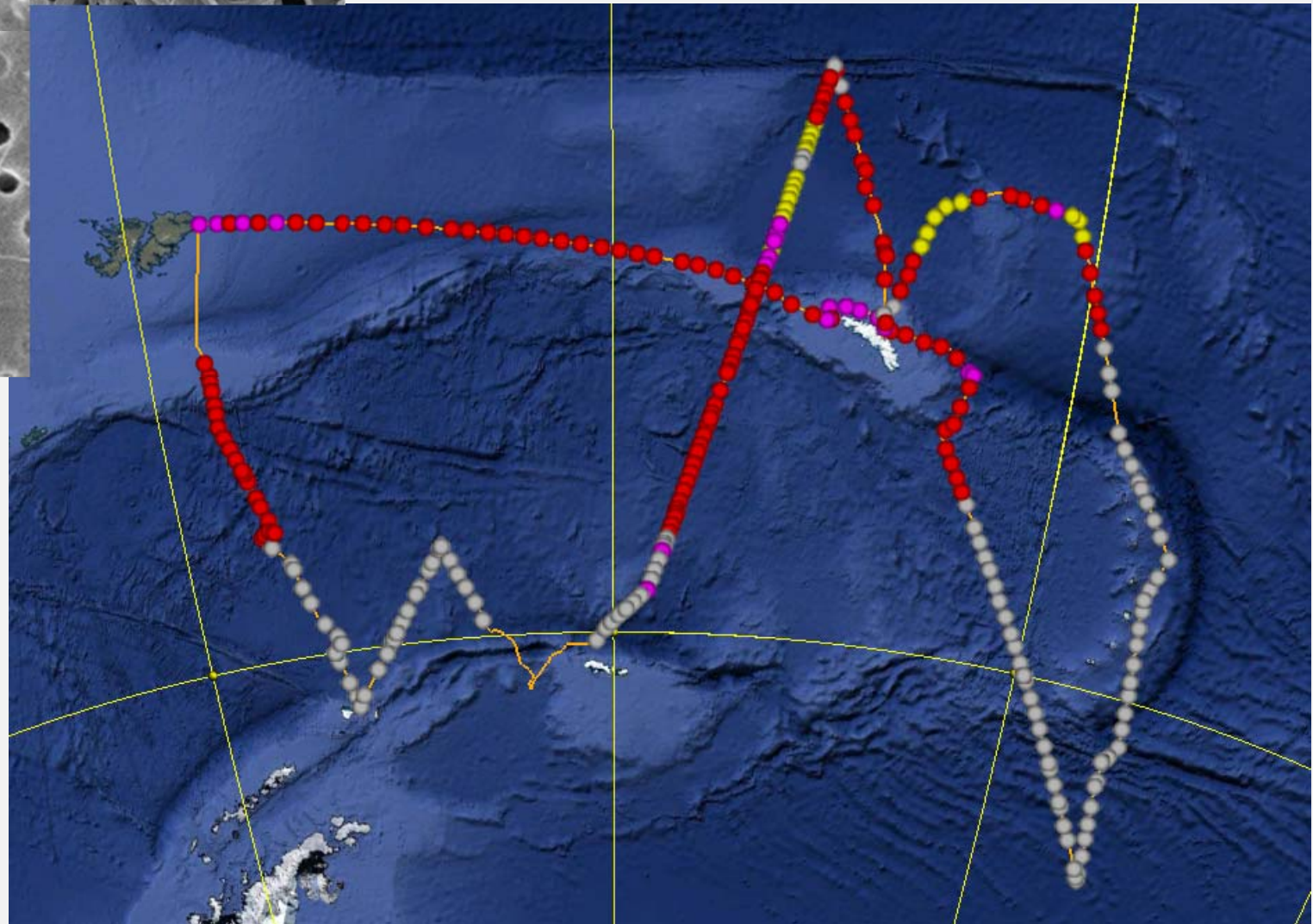
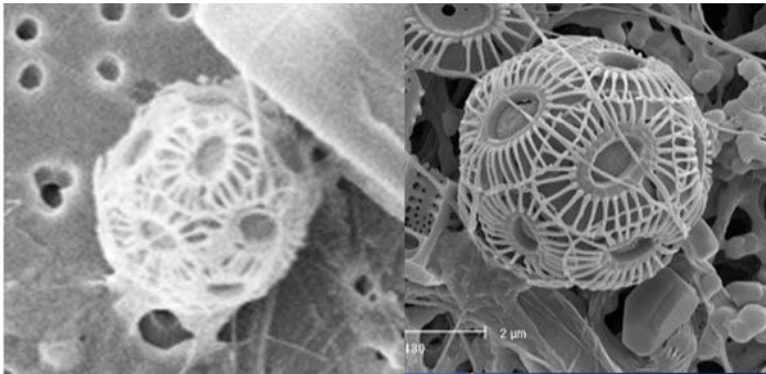


JR274 Antarctic  
Jan-Feb 2013

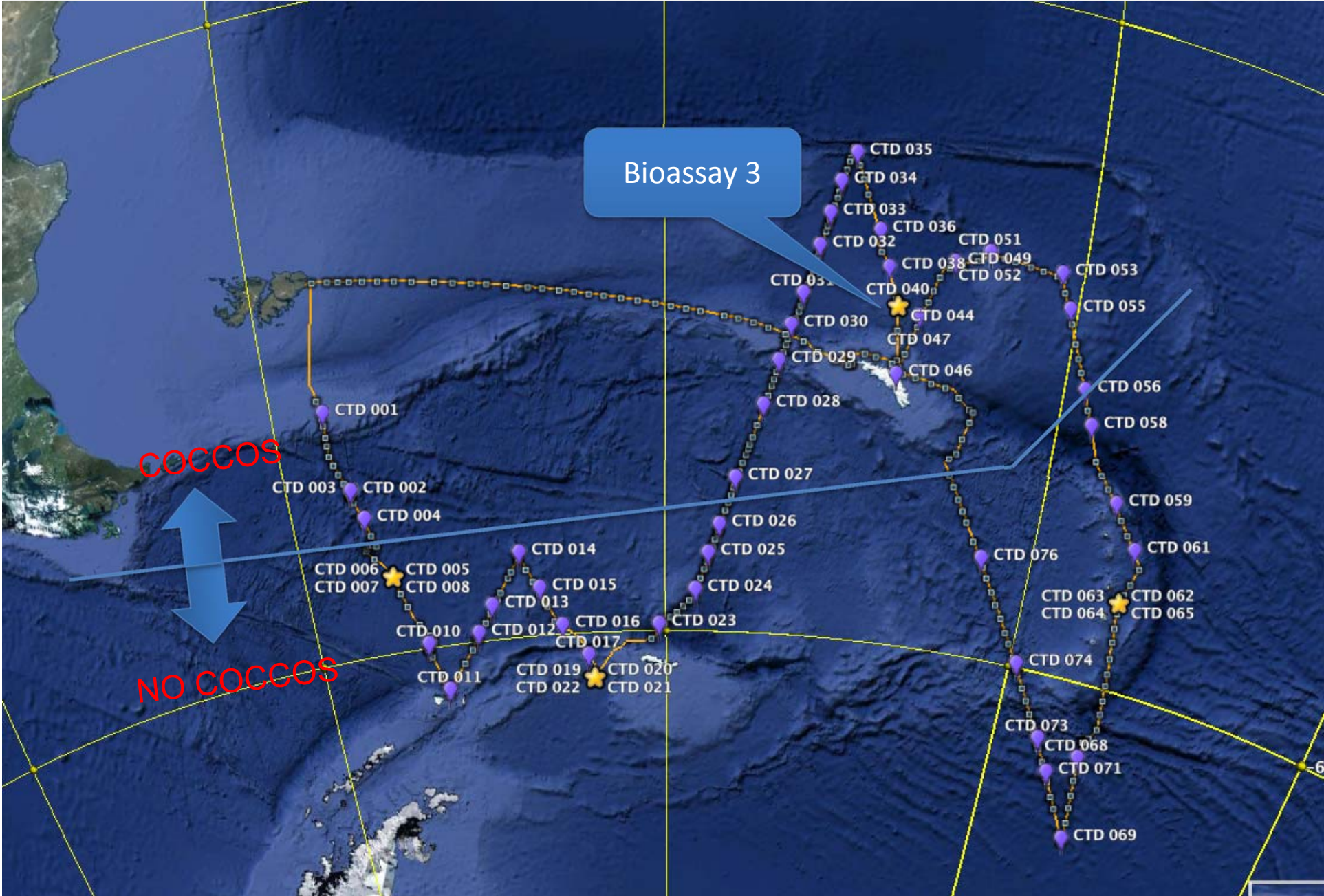


## *Emiliana huxleyi*

- Almost entirely type B/C (as predicted).
- Southerly limit rather sharp and crossed 4 times
- Some patchiness to N of South Georgia





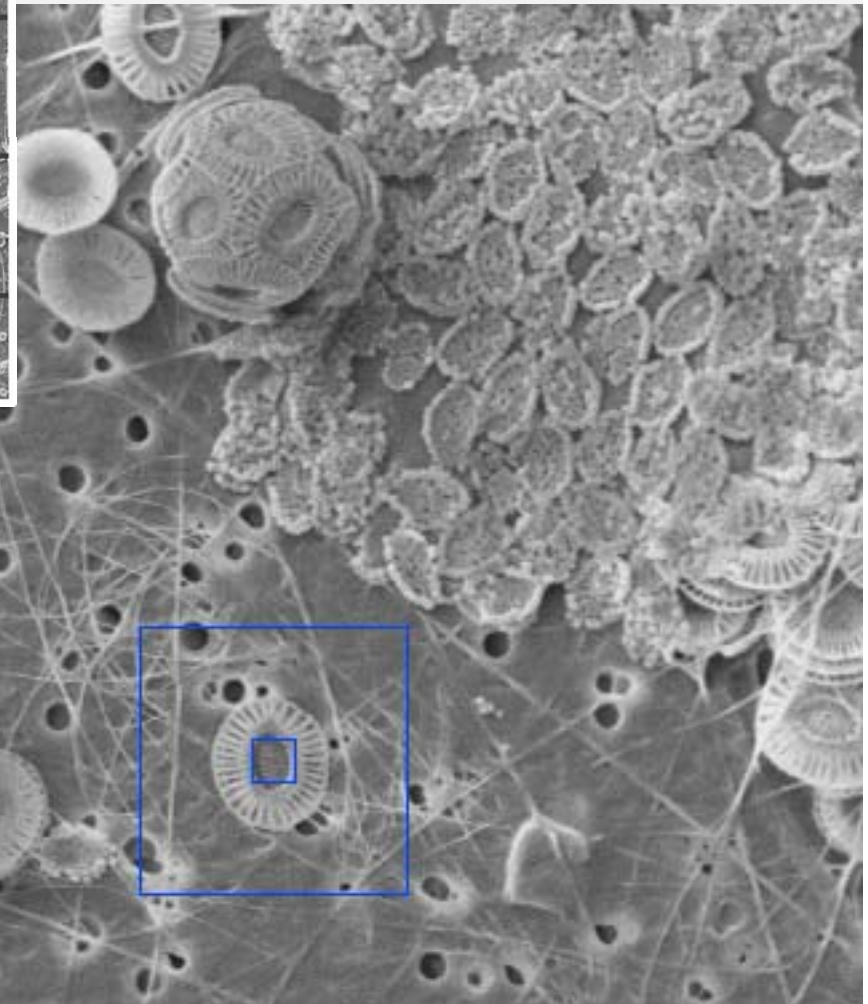
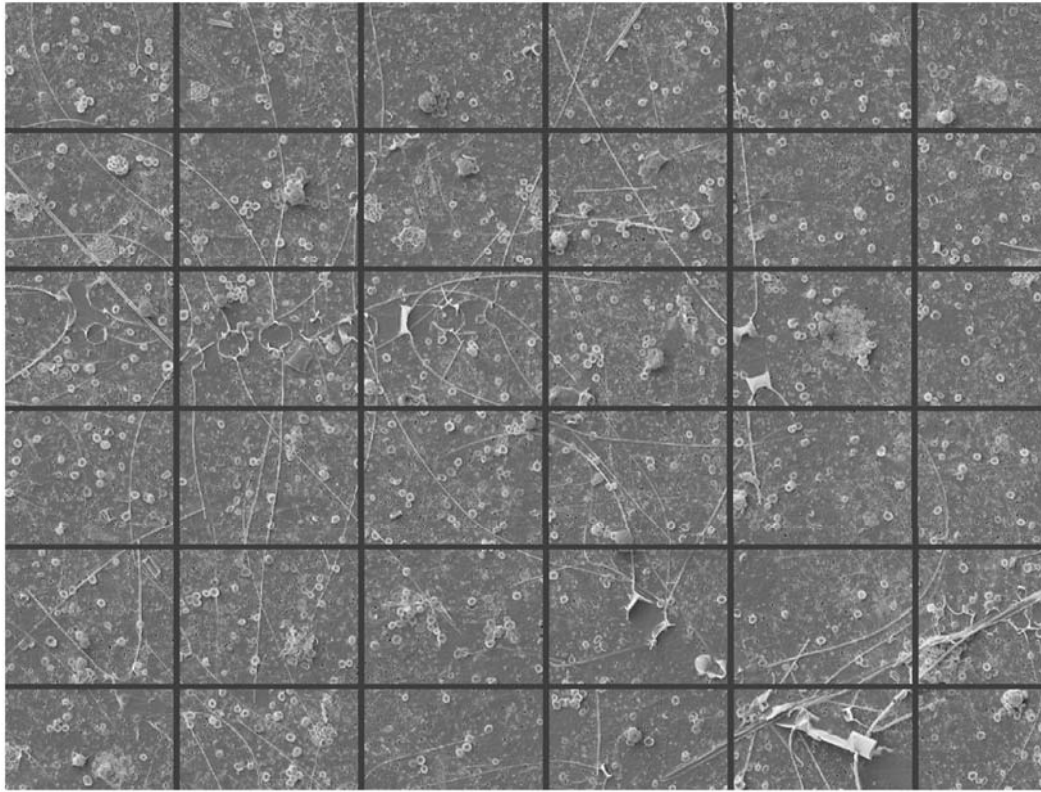


Bioassay 3

COCCOS

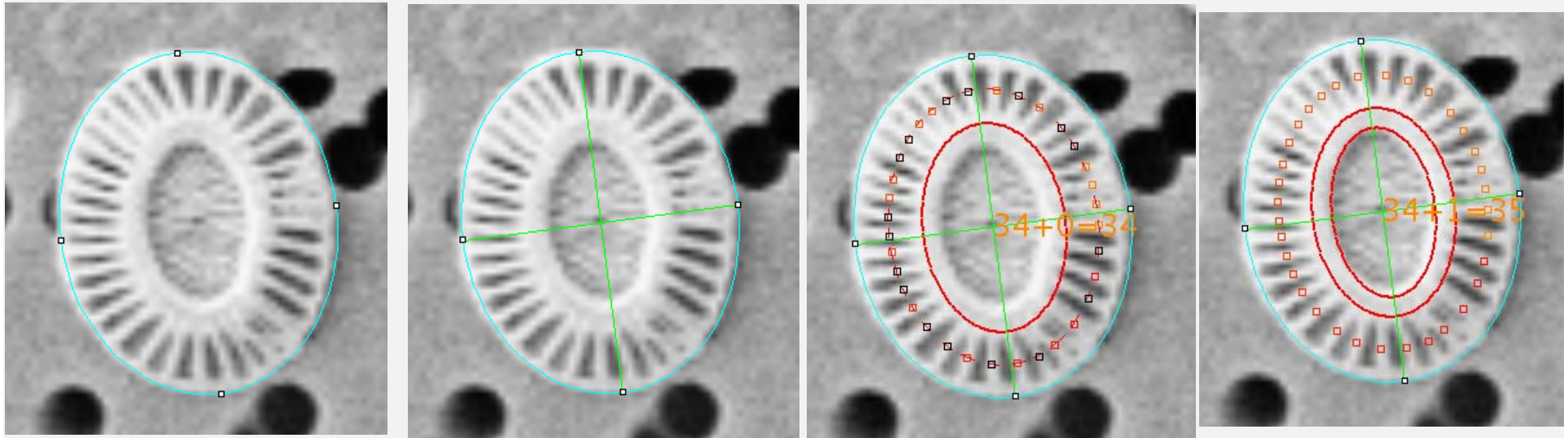
NO COCCOS

# image collection

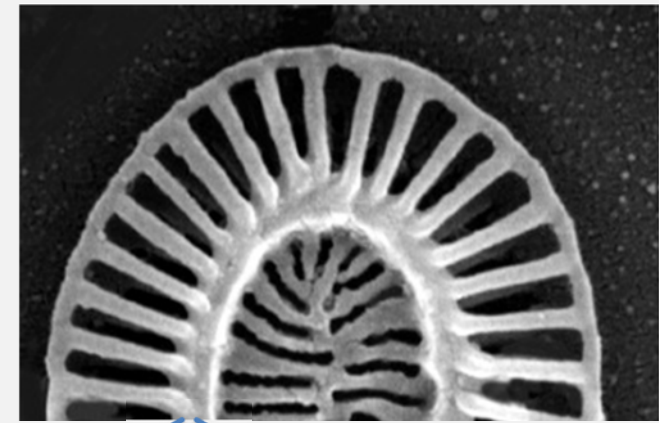


1. SEM automatically captures array of images overnight
2. Individual specimens are collected using imageJ macros

# semi-automated coccolith measurement using ImageJ/Fiji



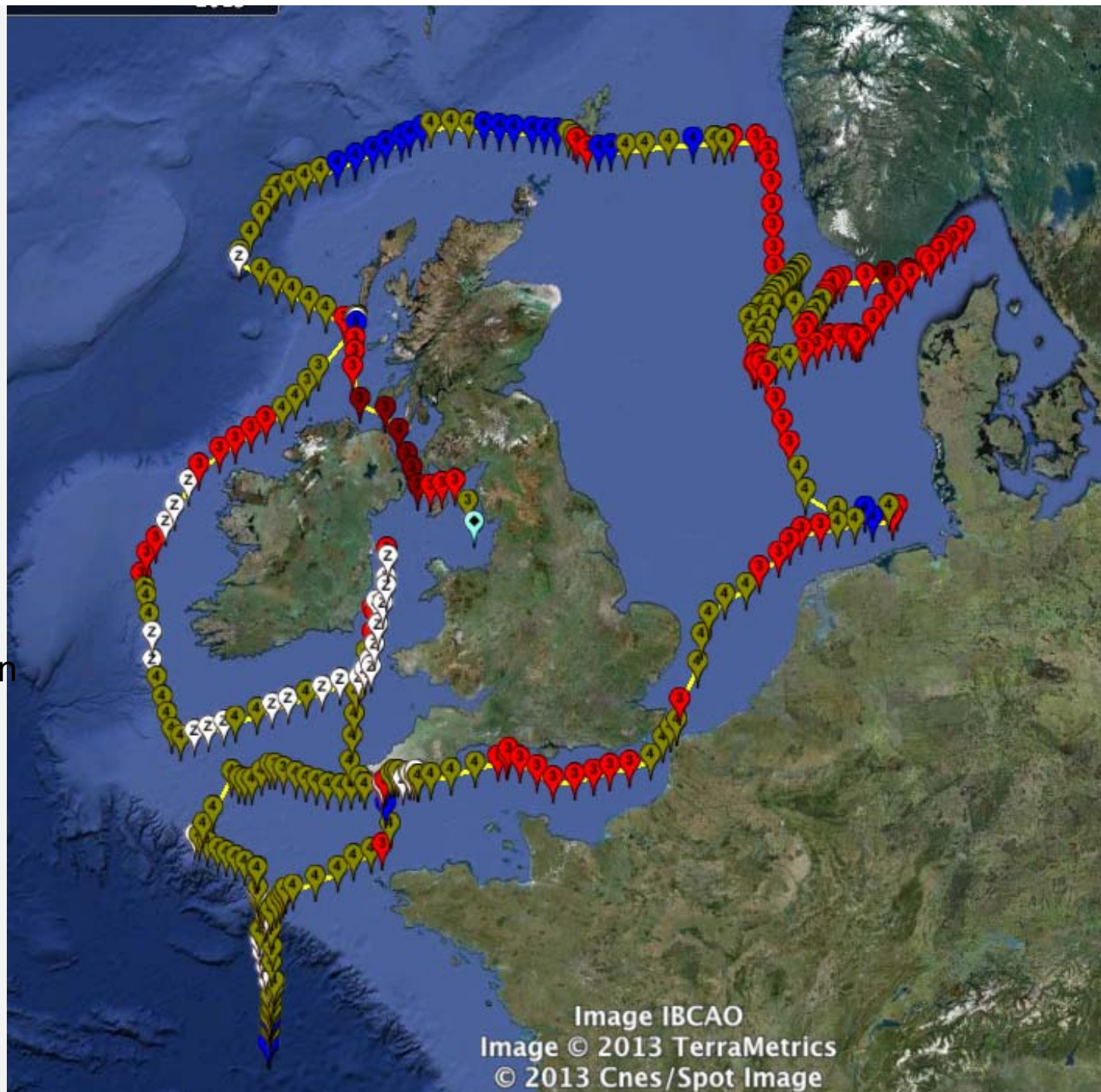
thickness of tube as proportion of coccolith width  
gives size-independent measure of degree of  
coccolith calcification

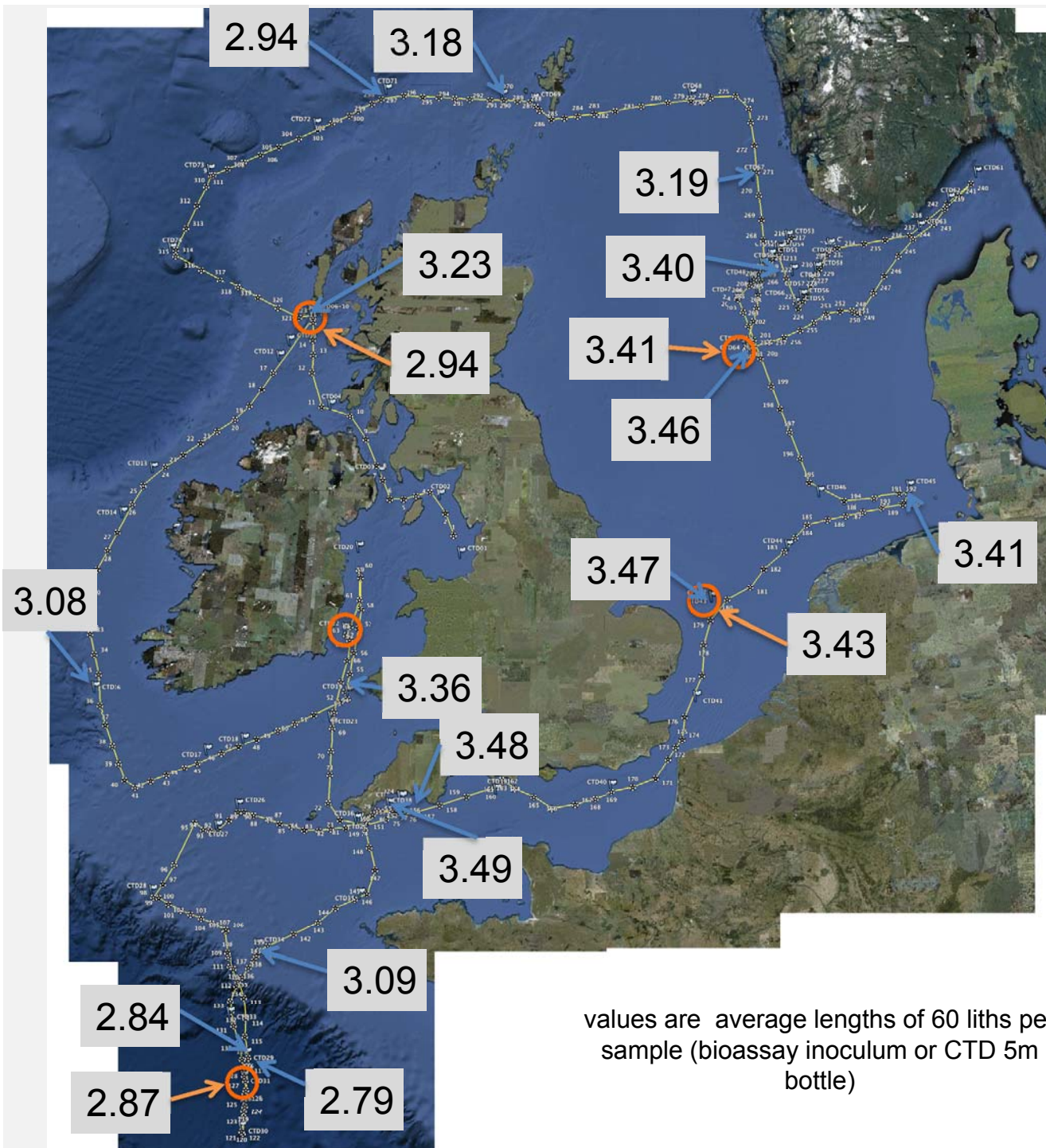


tt = tube thickness  
cw = coccolith width

Calcite saturation

- 4.5-5.0
- 4.0-4.5
- 3.5-4.0
- 3.0-3.5
- no data

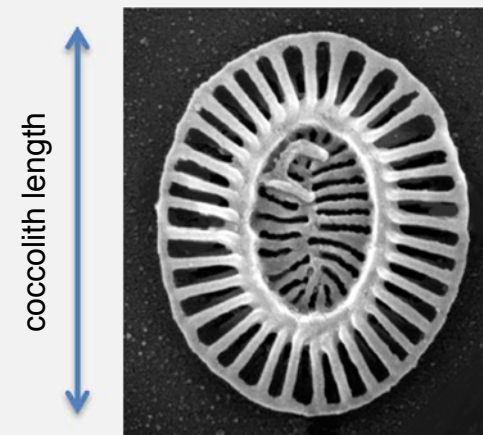


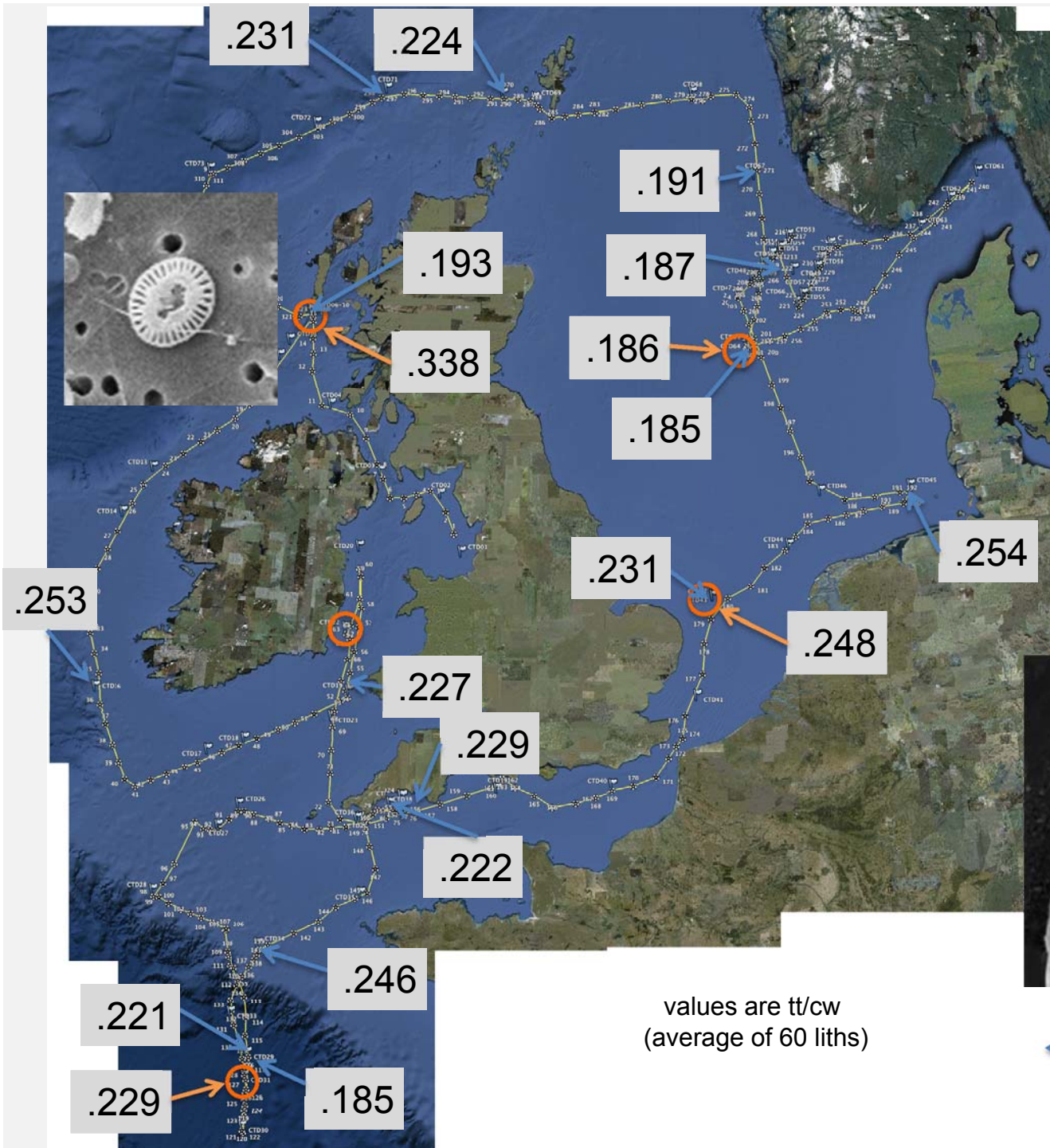


values are average lengths of 60 liths per sample (bioassay inoculum or CTD 5m bottle)

### *Emiliana huxleyi* length variation

larger specimens in more neritic samples (Channel and North Sea)

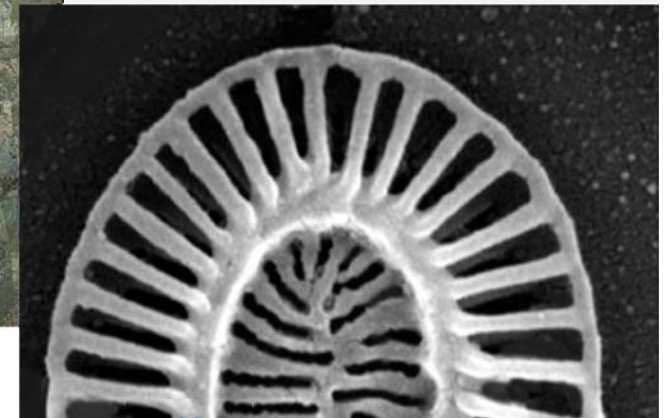




*Emiliana huxleyi*  
degree of  
calcification  
variation

total variation is  
low (0.18 to 0.25  
except for  
Bioassay 1), and  
no obvious  
pattern shown

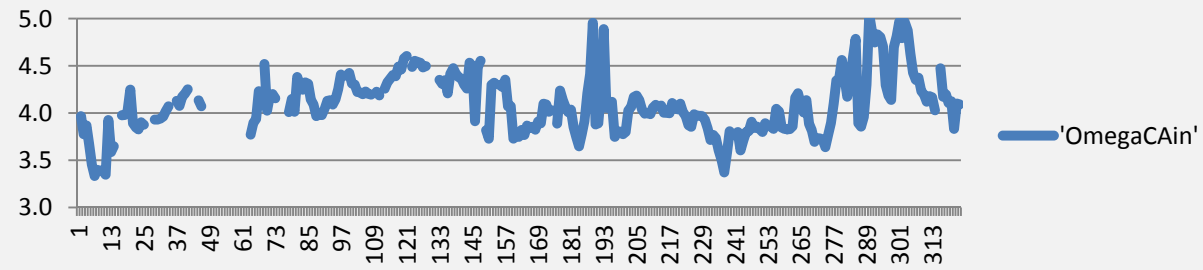
values are  $tt/cw$   
(average of 60 liths)



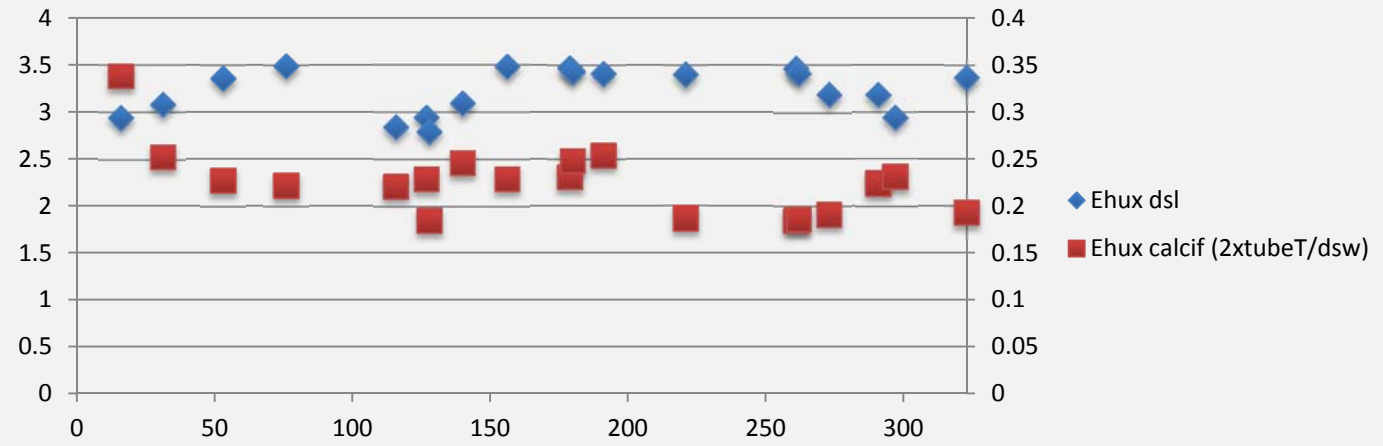
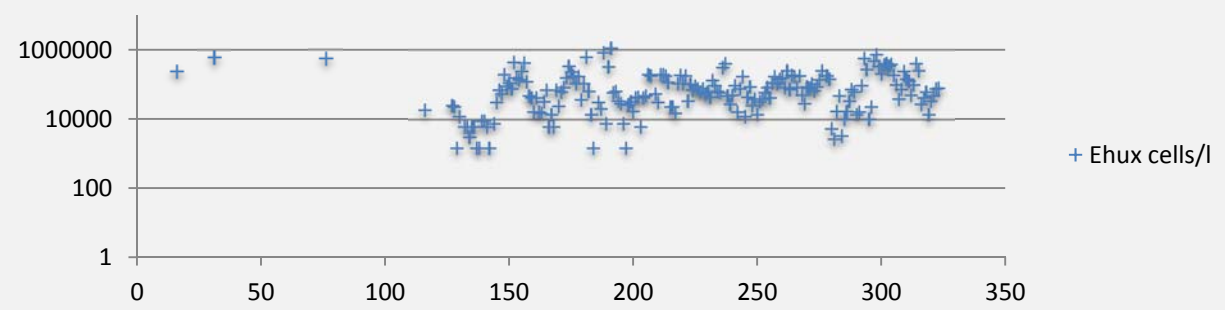
$tt$  = tube thickness

$cw$  = coccolith width

### 'OmegaCAin'



### Ehux cells/l



No obvious first order relationship of *E. huxleyi* abundance, size or degree of calcification with calcite saturation state

# Bioassays

5 incubations each 4 days at four CO<sub>2</sub> levels  
(380, 500, 750, 1000ppm)

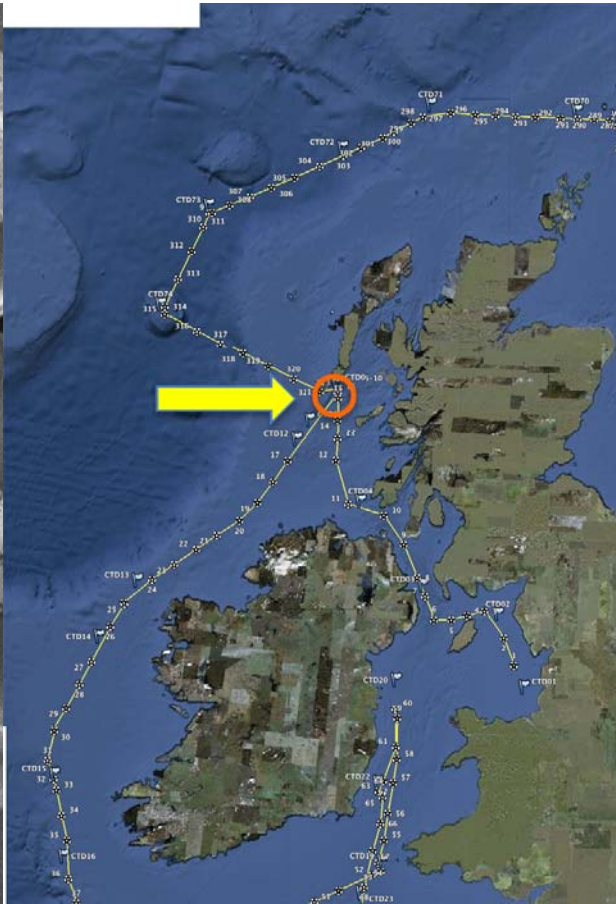
very extensive matrix of observations



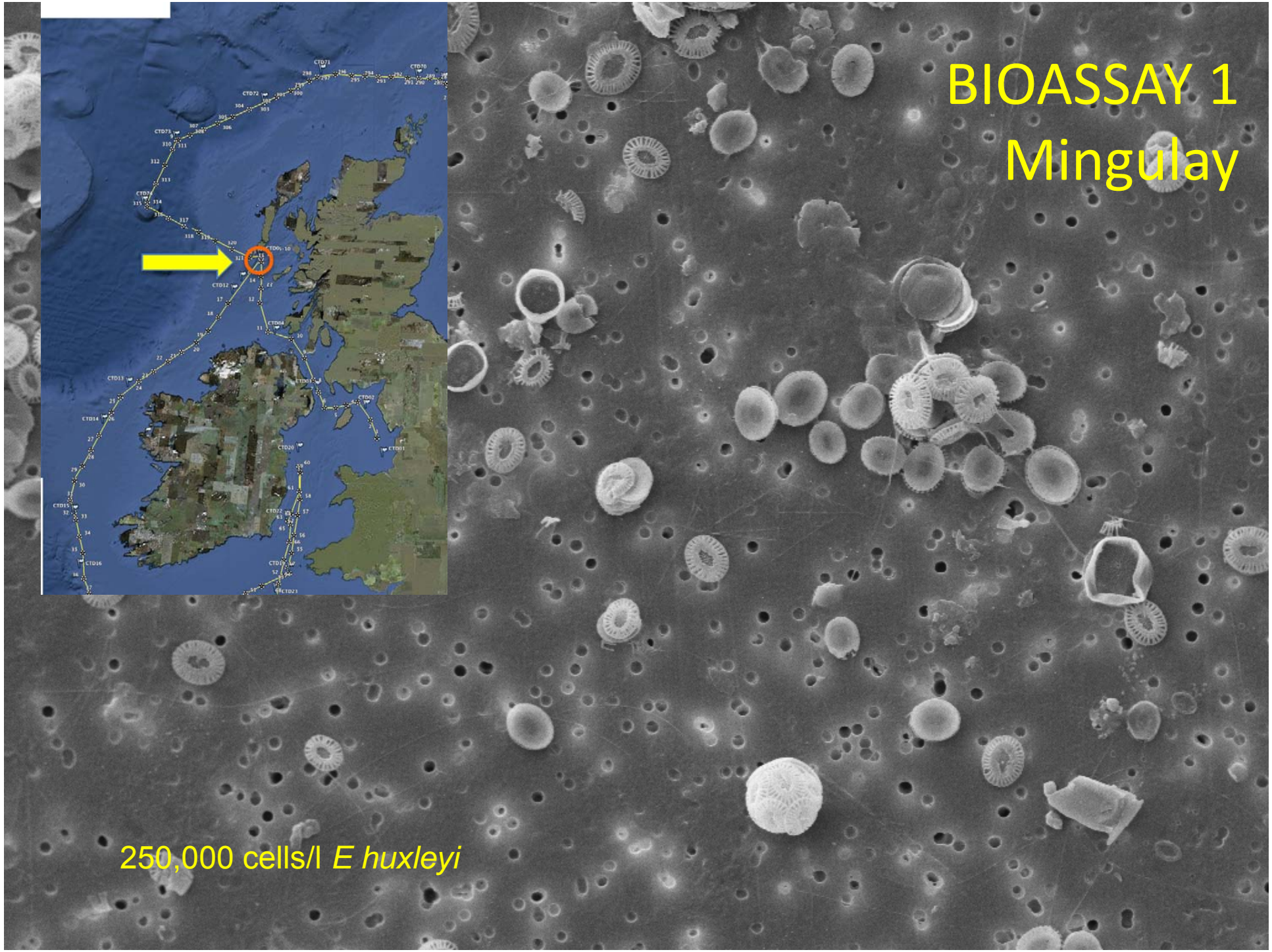


# BIOASSAY 1

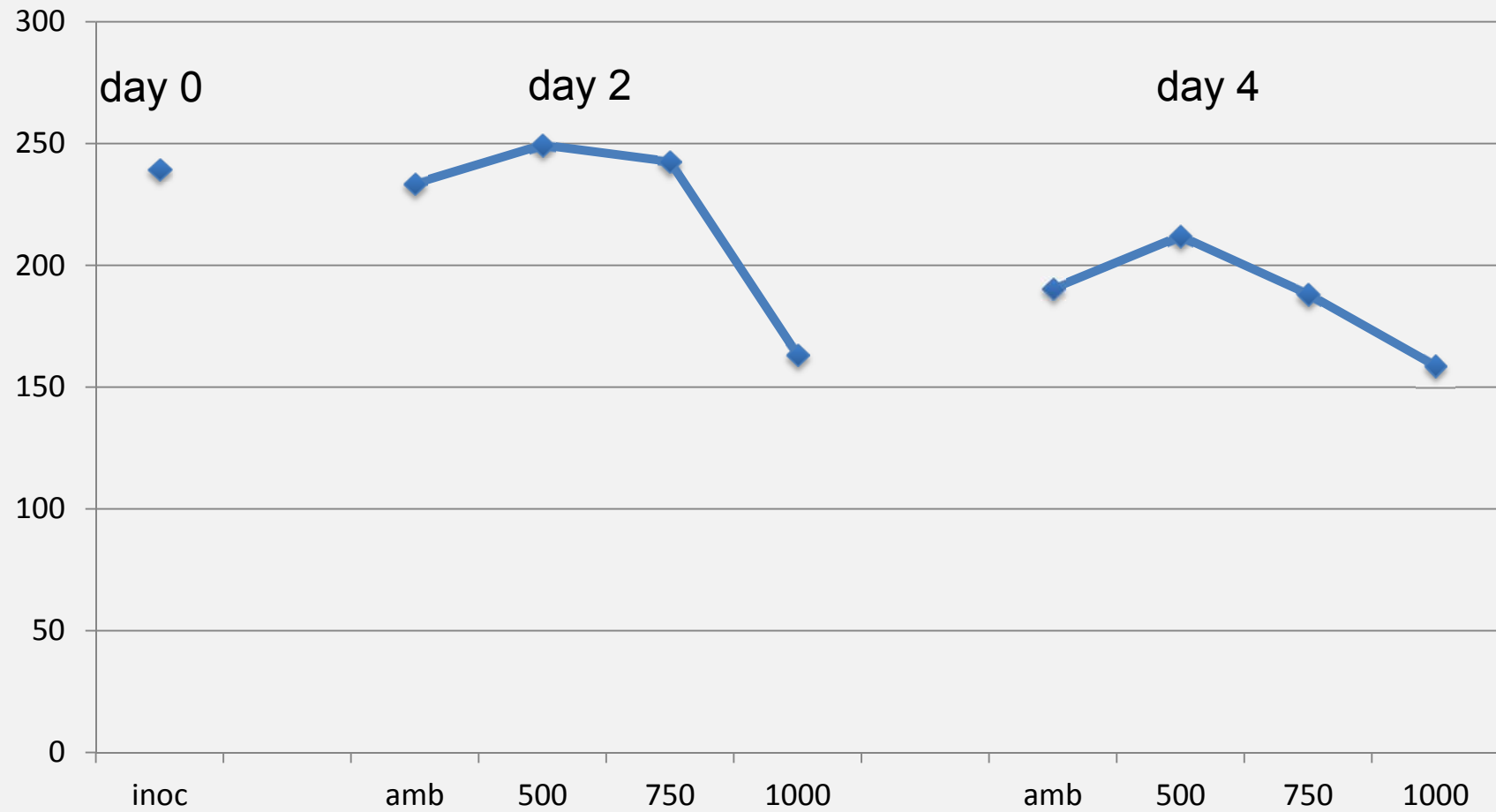
## Mingulay



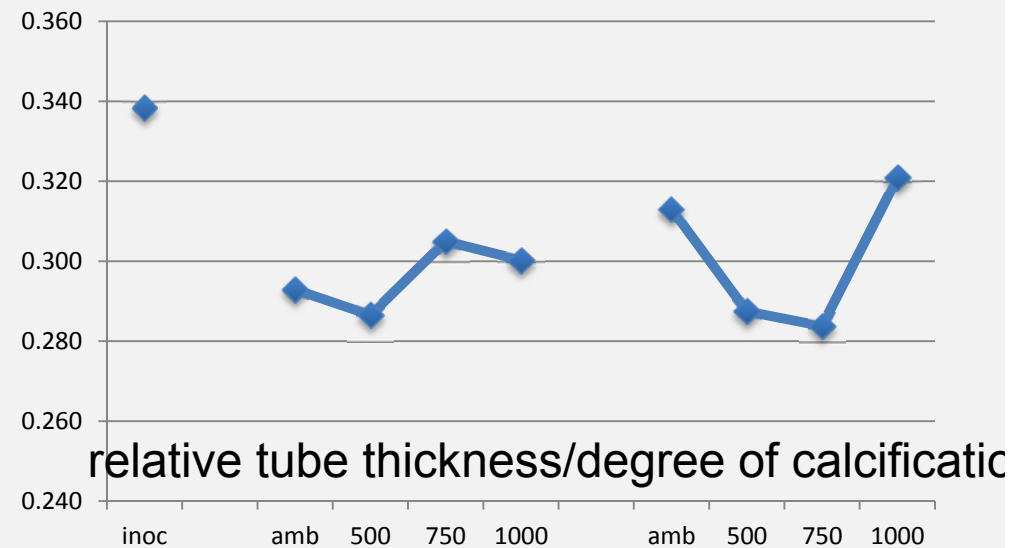
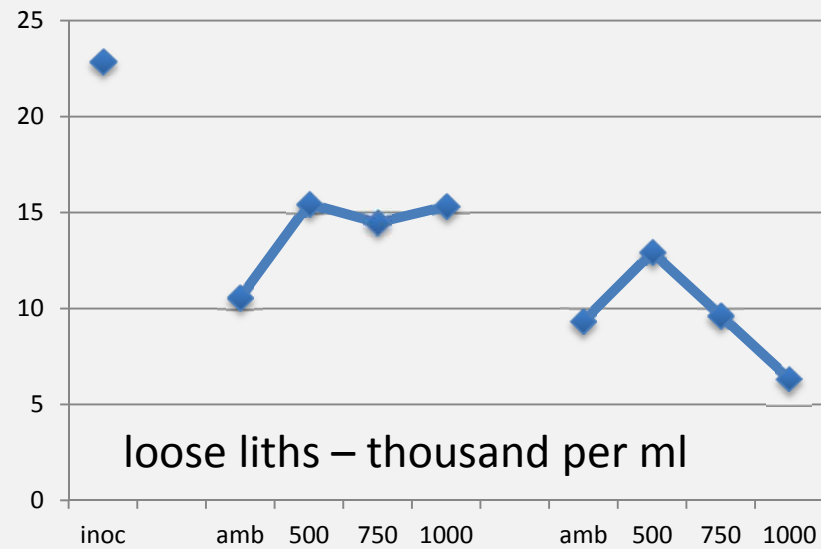
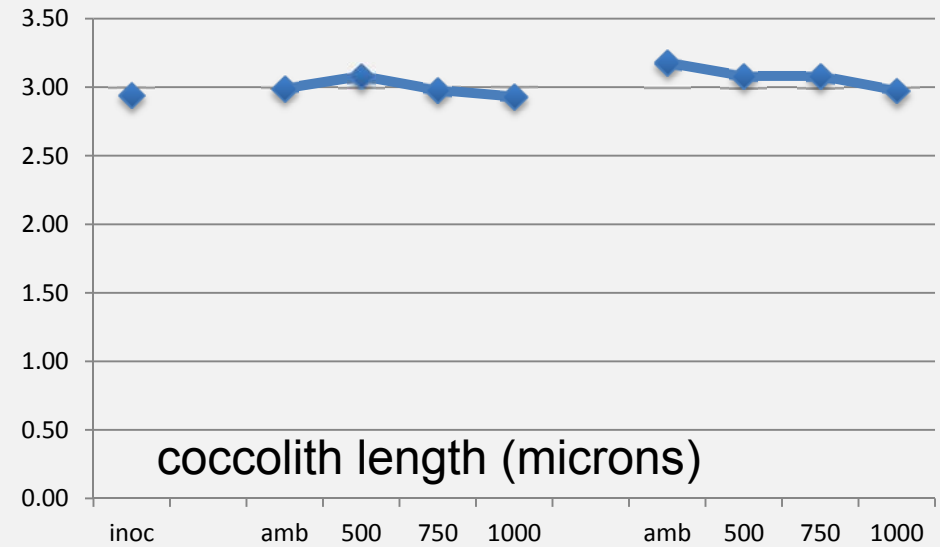
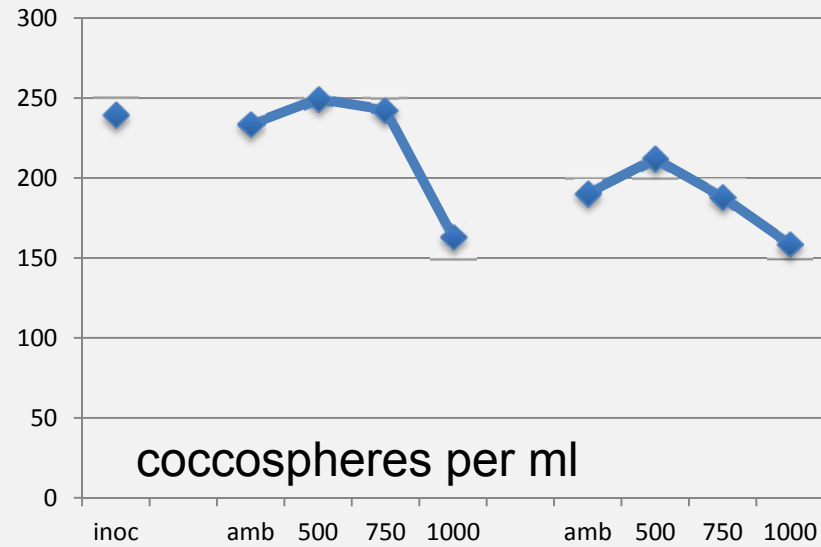
250,000 cells/l *E huxleyi*



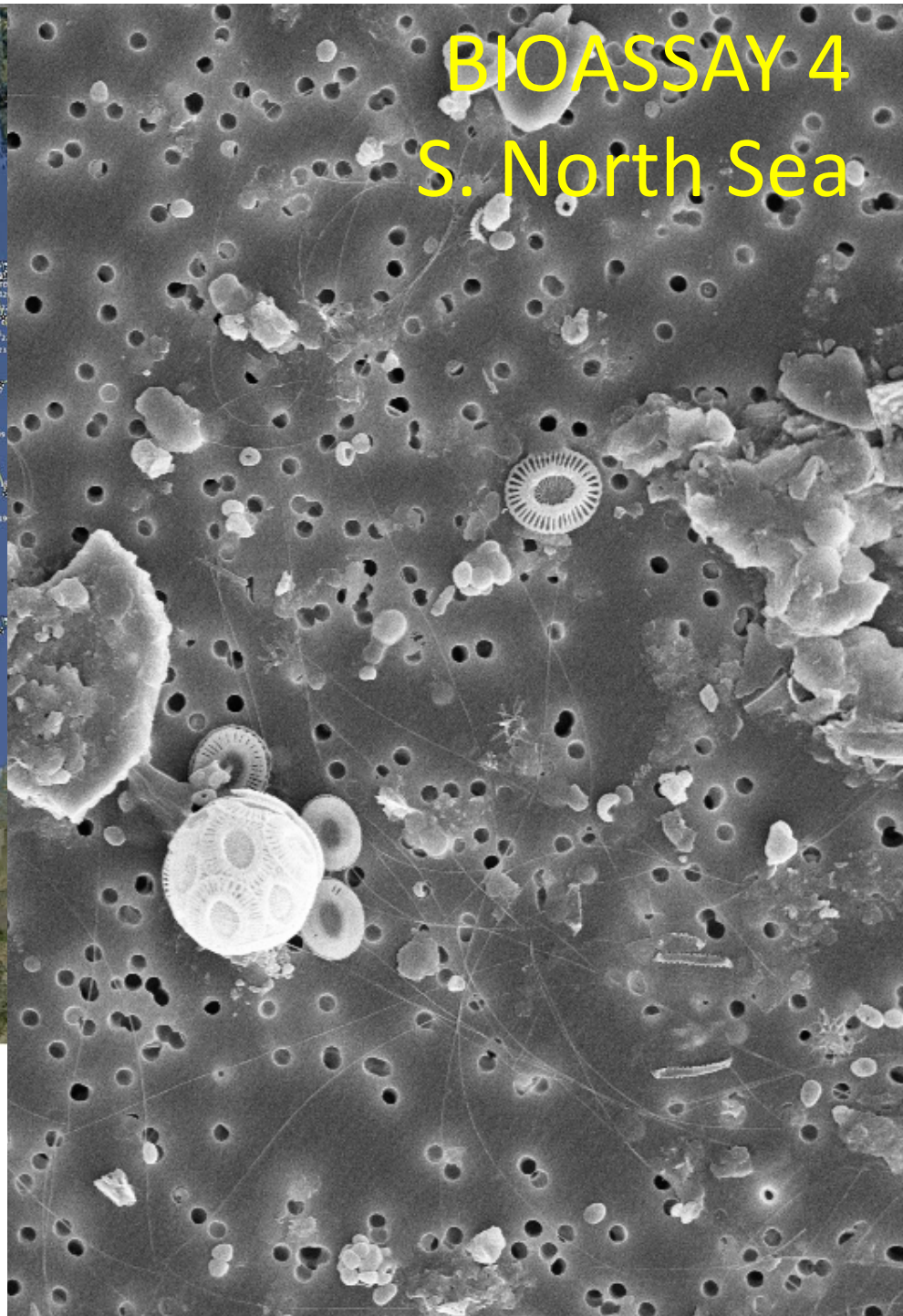
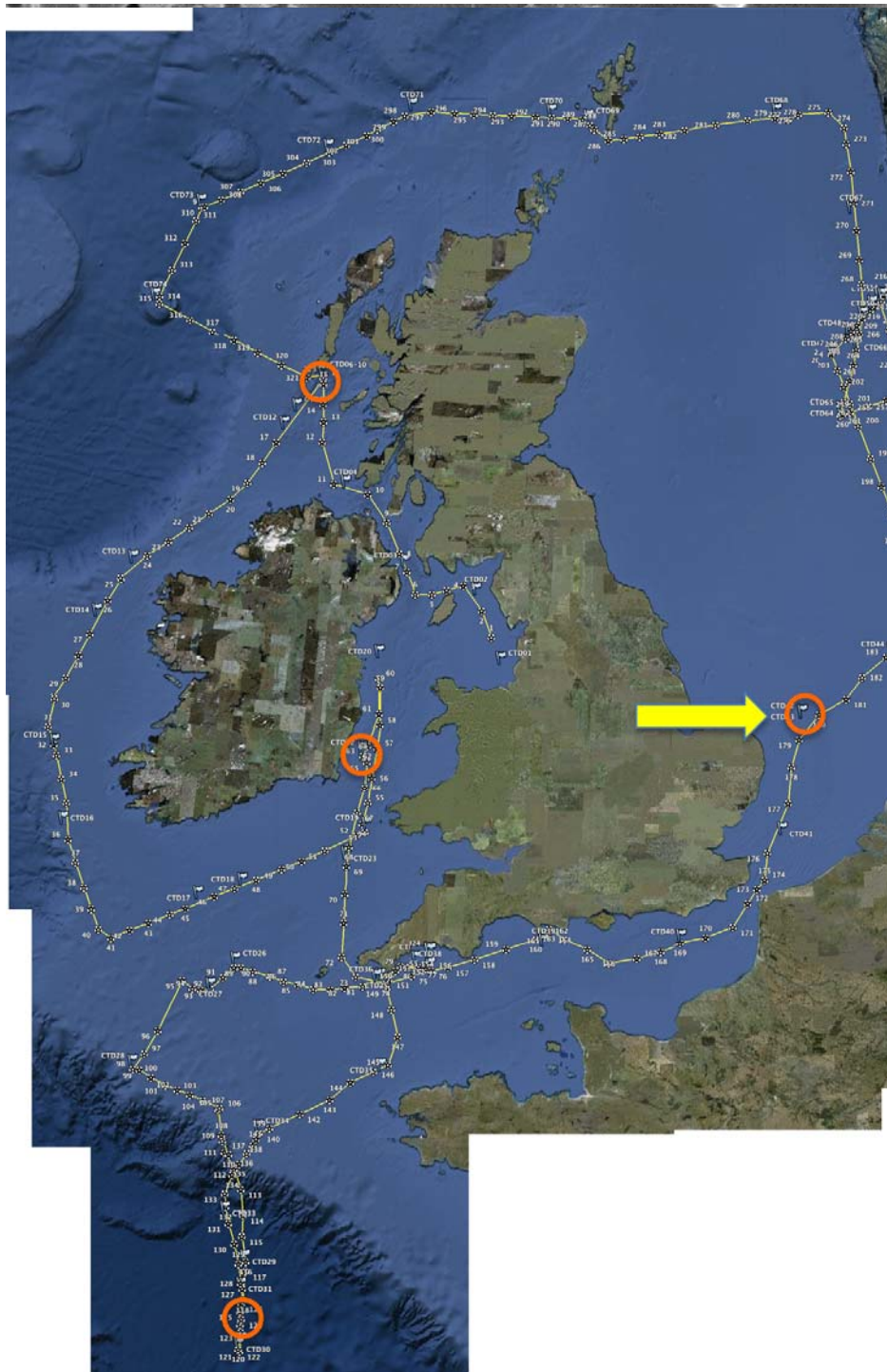
# coccospheres/ml



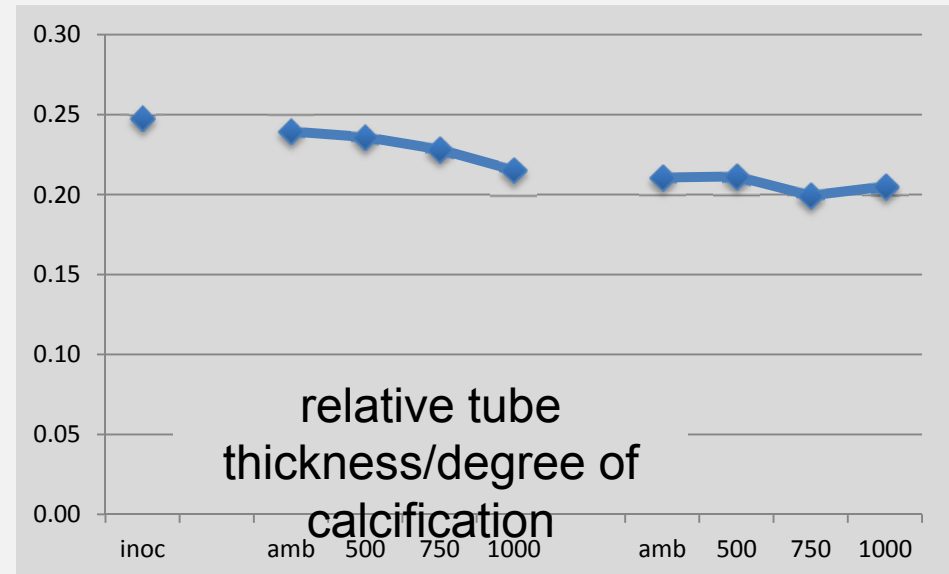
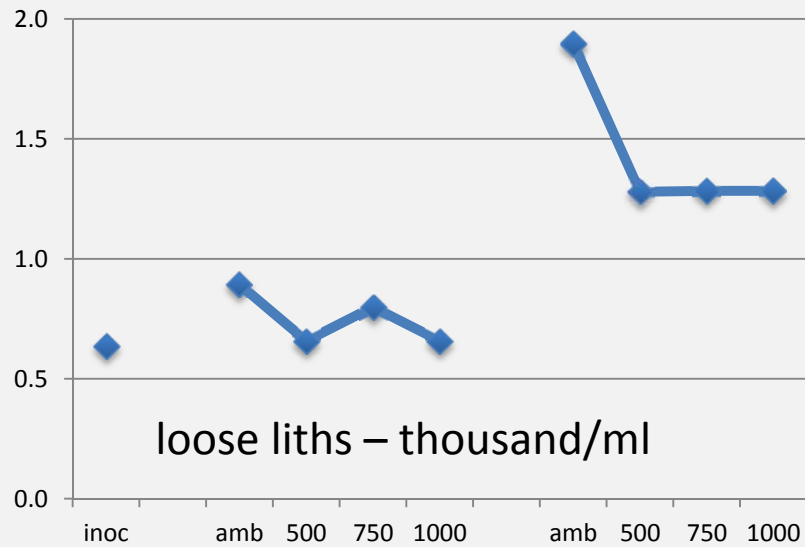
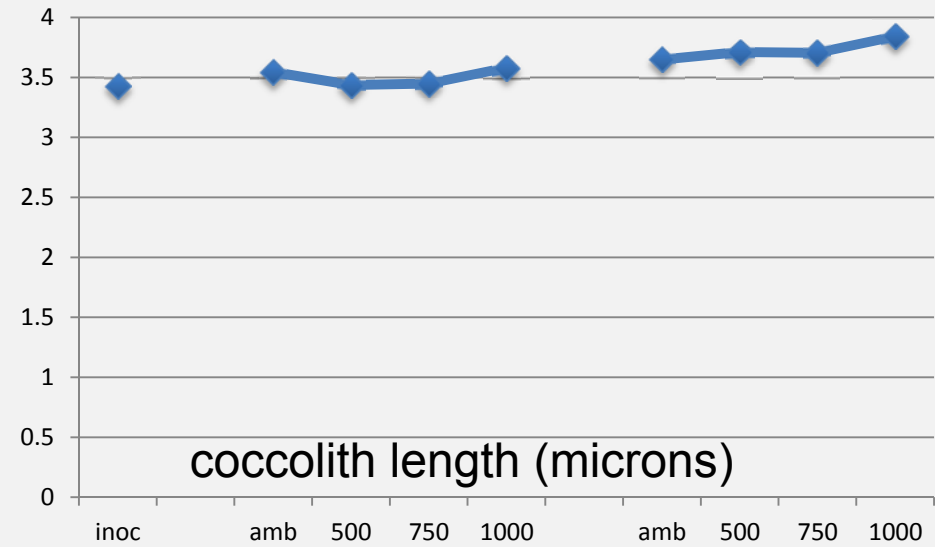
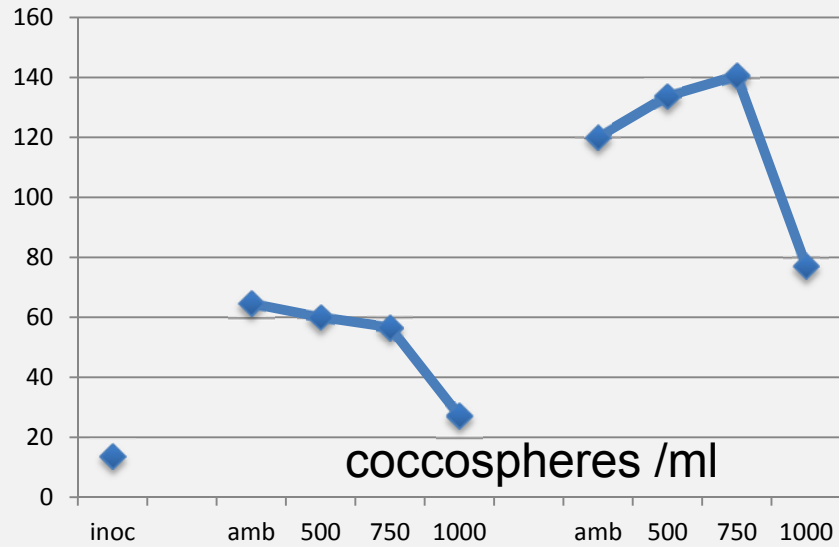
# Bioassay 1 - Mingulay



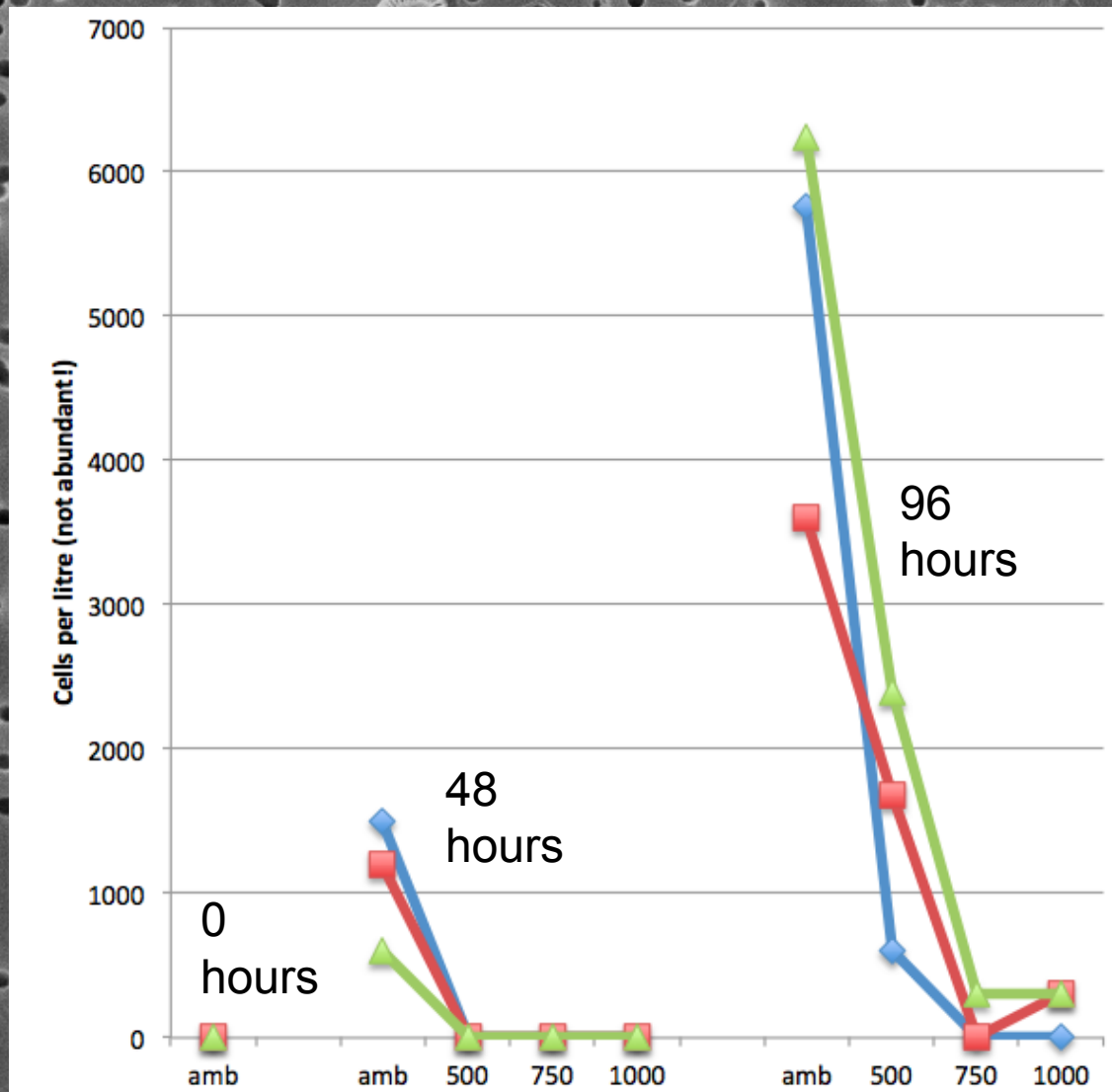
# BIOASSAY 4 S. North Sea



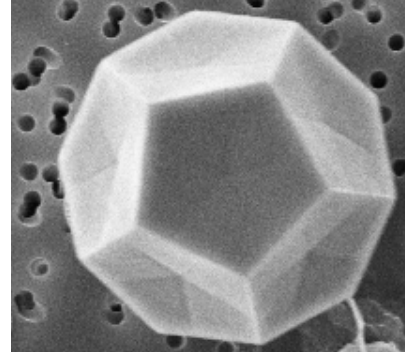
# Bioassay 4 – S. North Sea



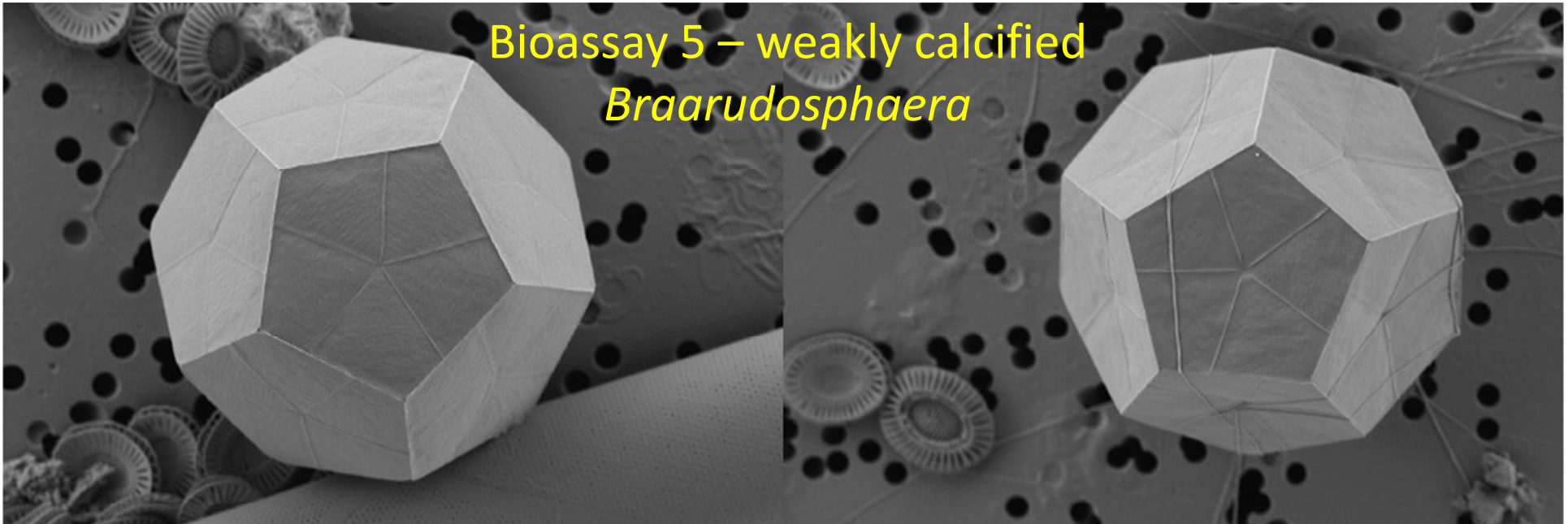
# D366 - Bioassay 5 - North Sea



*Braarudosphaera bigelowii*  
- growing population (low ab)  
- only in low CO<sub>2</sub> treatments

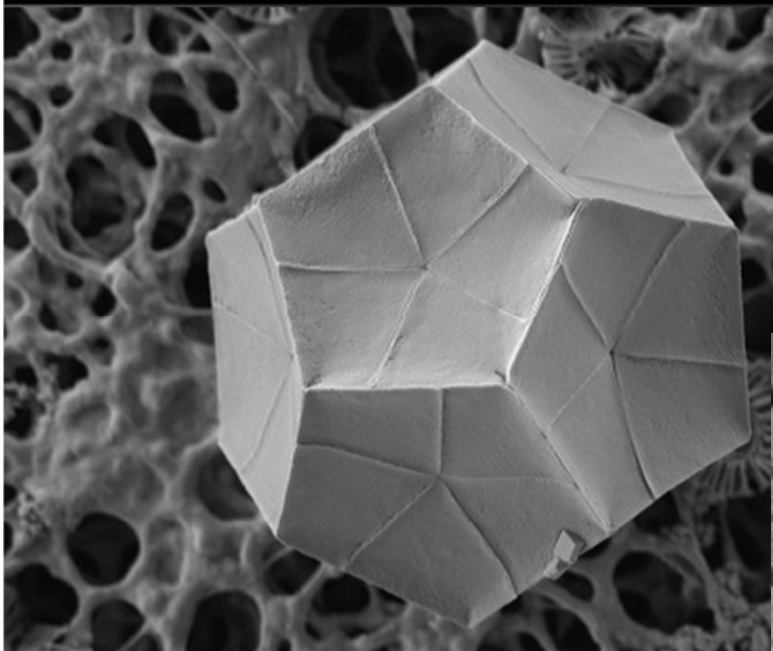


# Bioassay 5 – weakly calcified *Braarudosphaera*

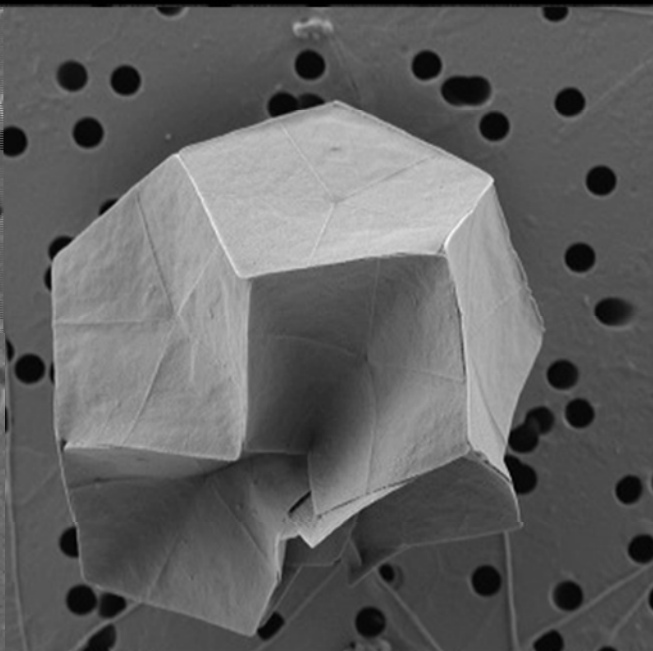
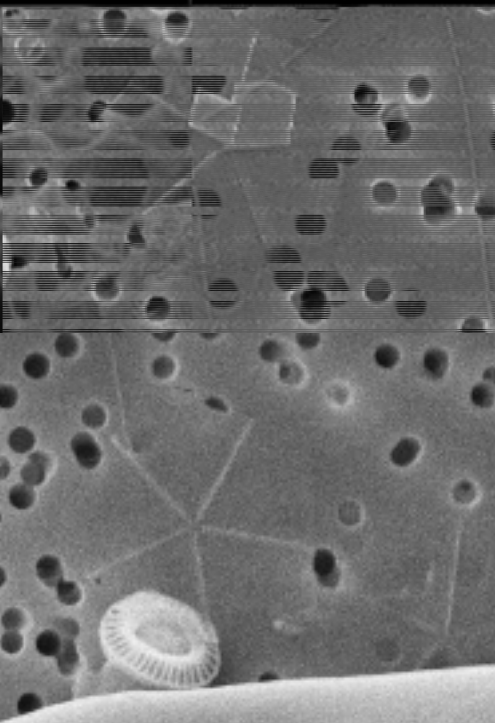


1 μm  
Mag = 13.66 KX 3.00 KV Signal A = SE2  
Image Pixel Size = 10.7 nm Signal B = ESB  
WD = 5.4 mm Signal = 0.4145  
Wednesday, October 31, 2012 Aperture Size = 30.00 μm  
1.74e-004 Pa  
File Name = JF924-005.91 N = 4

2 μm  
Mag = 12.74 KX 3.00 KV Signal A = SE2  
Image Pixel Size = 11.5 nm Signal B = ESB  
WD = 5.3 mm Signal = 0.4145  
Wednesday, October 31, 2012 Aperture Size = 30.00 μm  
1.66e-004 Pa  
File Name = JF924-007.91 N = 4

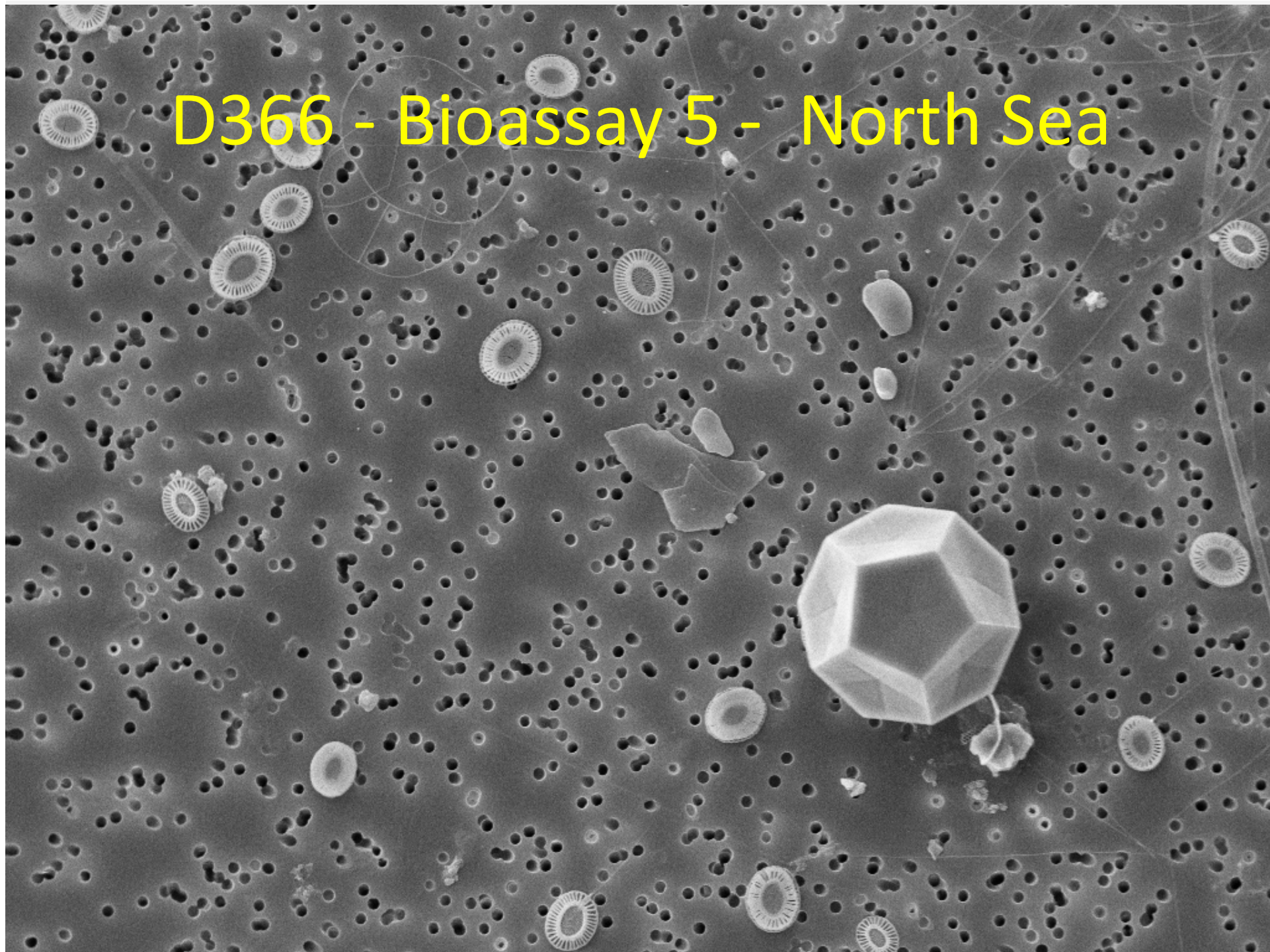


1 μm  
Mag = 15.51 KX 3.00 KV Signal A = SE2  
Image Pixel Size = 9.4 nm Signal B = ESB  
WD = 5.2 mm Signal = 0.4145  
Wednesday, October 31, 2012 Aperture Size = 30.00 μm  
1.54e-004 Pa  
File Name = JF924-008.91 N = 4



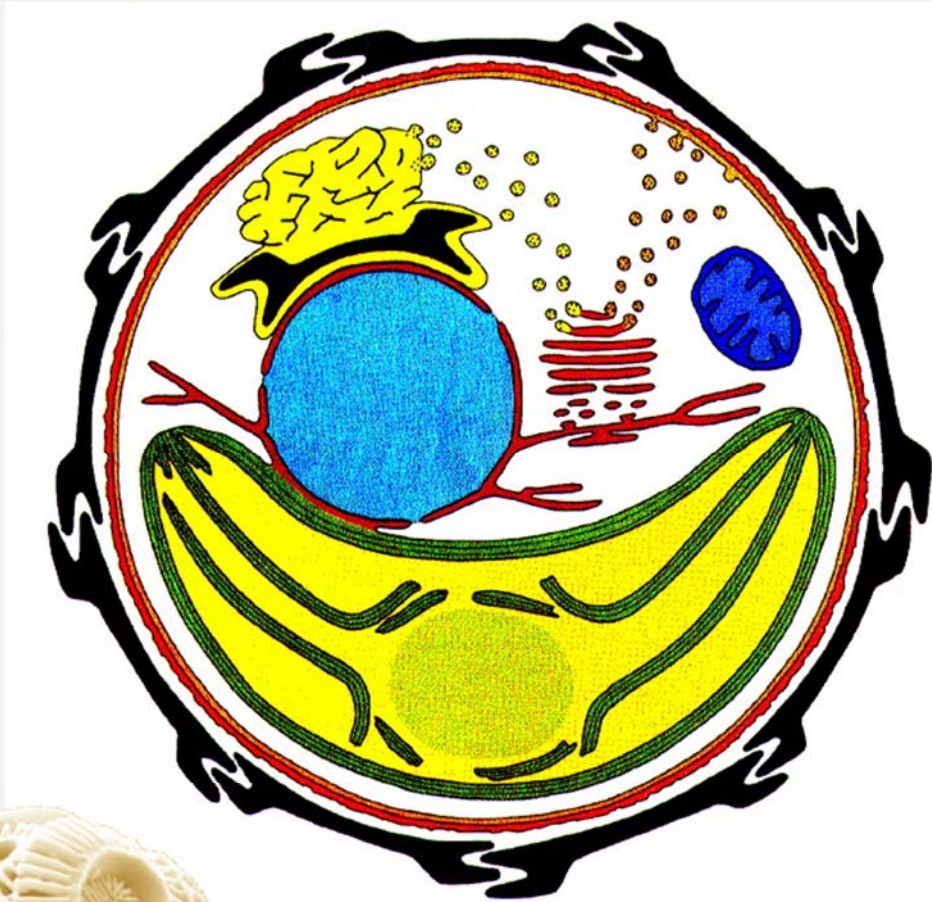
1 μm  
Mag = 15.51 KX 3.00 KV Signal A = SE2  
Image Pixel Size = 9.4 nm Signal B = ESB  
WD = 5.2 mm Signal = 0.4145  
Wednesday, October 31, 2012 Aperture Size = 30.00 μm  
1.54e-004 Pa  
File Name = JF924-008.91 N = 4

# D366 - Bioassay 5 - North Sea

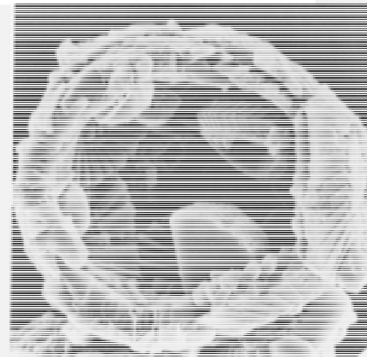




# intracellular calcification

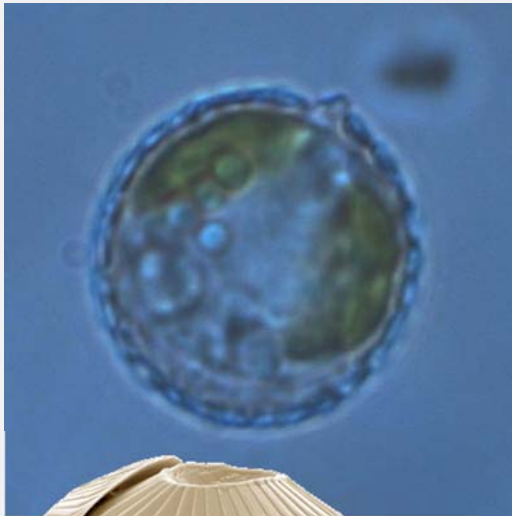


- Calcification inside vesicle deep within cell
- Transport of  $\text{Ca}^{2+}$  to vesicle
- Calcification from vesicle fluid not seawater
- Trace-element chemistry highly controlled
- Muted response to OA
- Shown by modern and fossil heterococcoliths

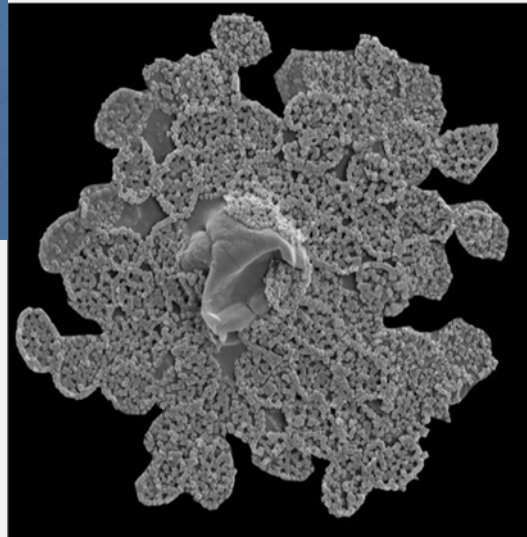


# extra-cellular calcification

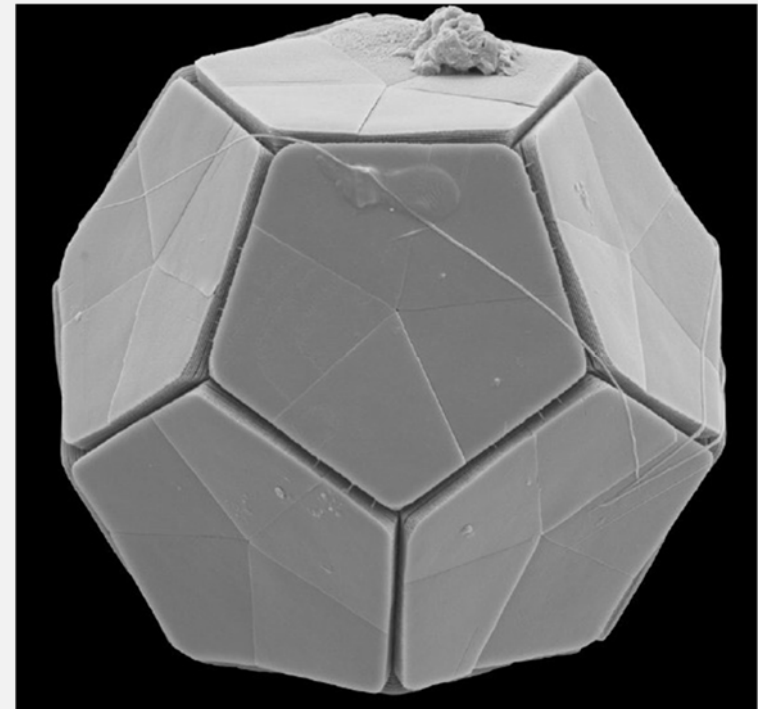
Holococcoliths,  
observational evidence



*Coccolithus pelagicus* - *Holococcoliths*



*Braarudosphaera*,  
morphologically unfeasible

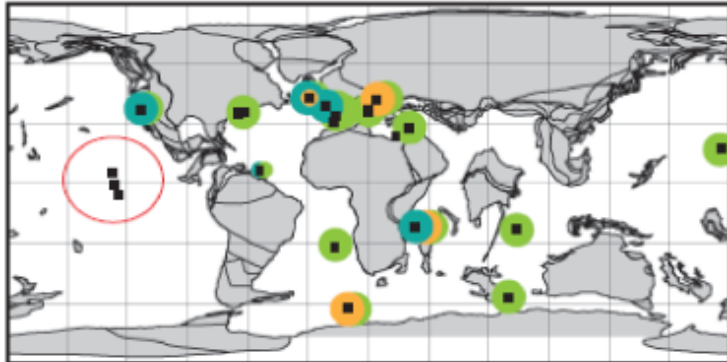


*Braarudosphaera bigelowii*

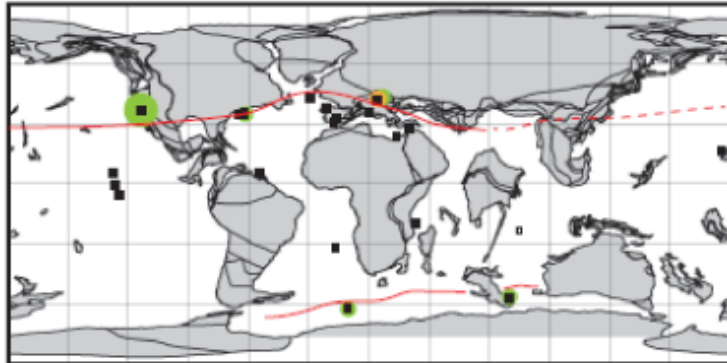
# Geological evidence

- work in prep of Sam Gibbs & Paul Bown
- Holococcoliths and *Braarudosphaera* seem to be especially heavily affected by the Paleocene Eocene Thermal Maximum OA event

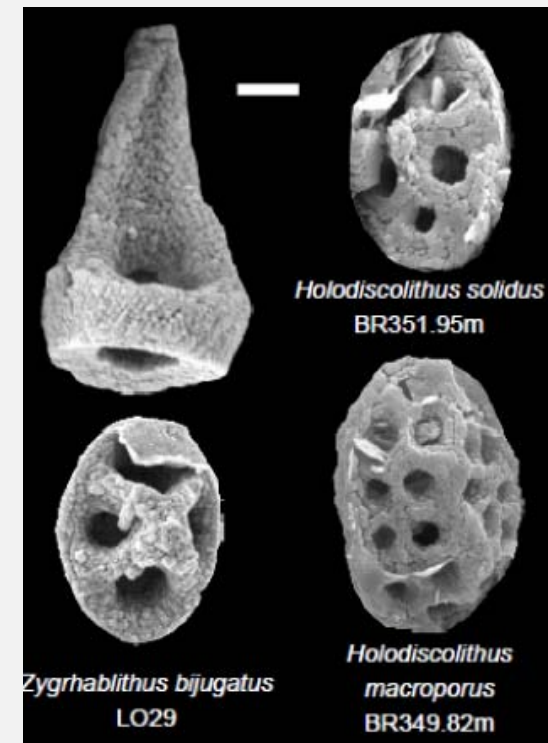
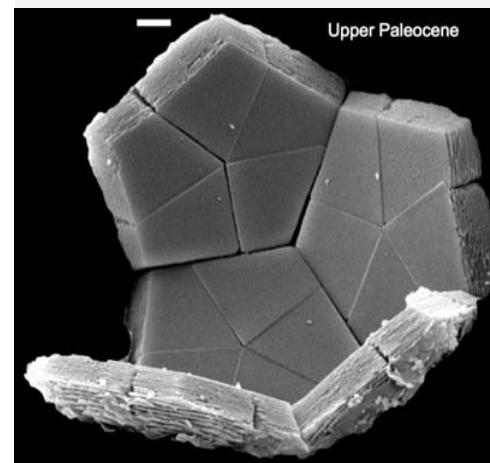
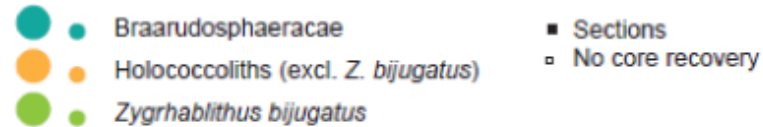
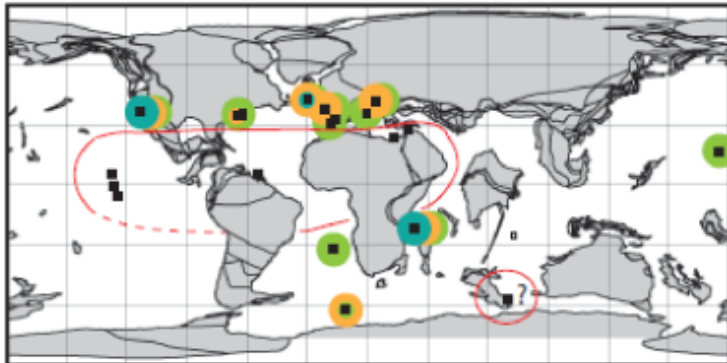
Post-PETM



PETM



Pre-PETM



# Summary

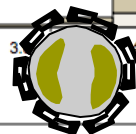
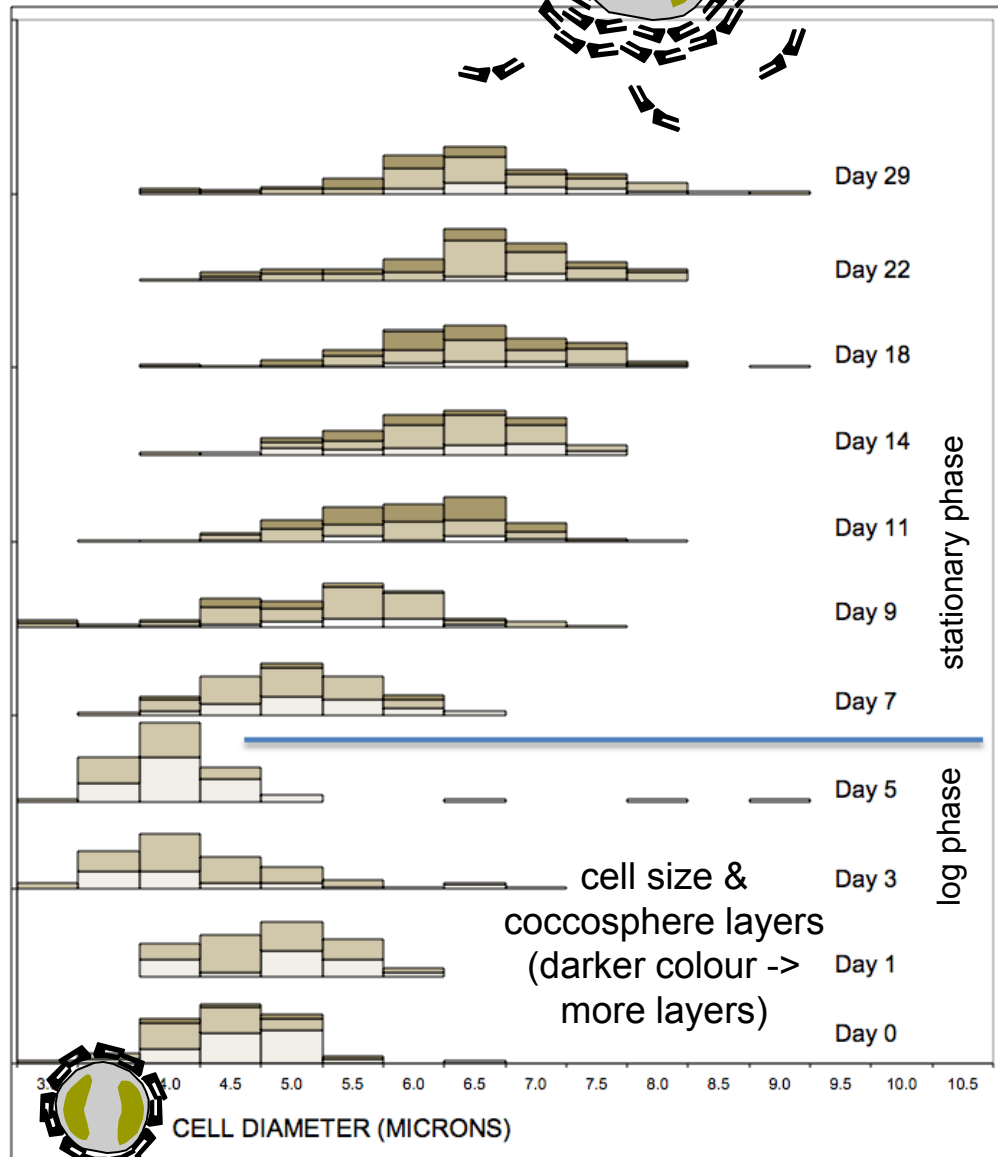
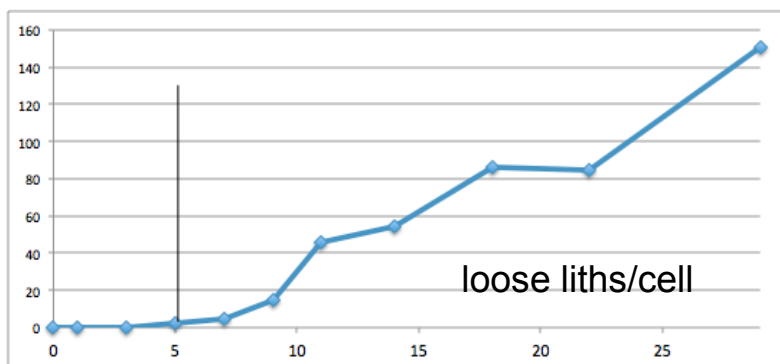
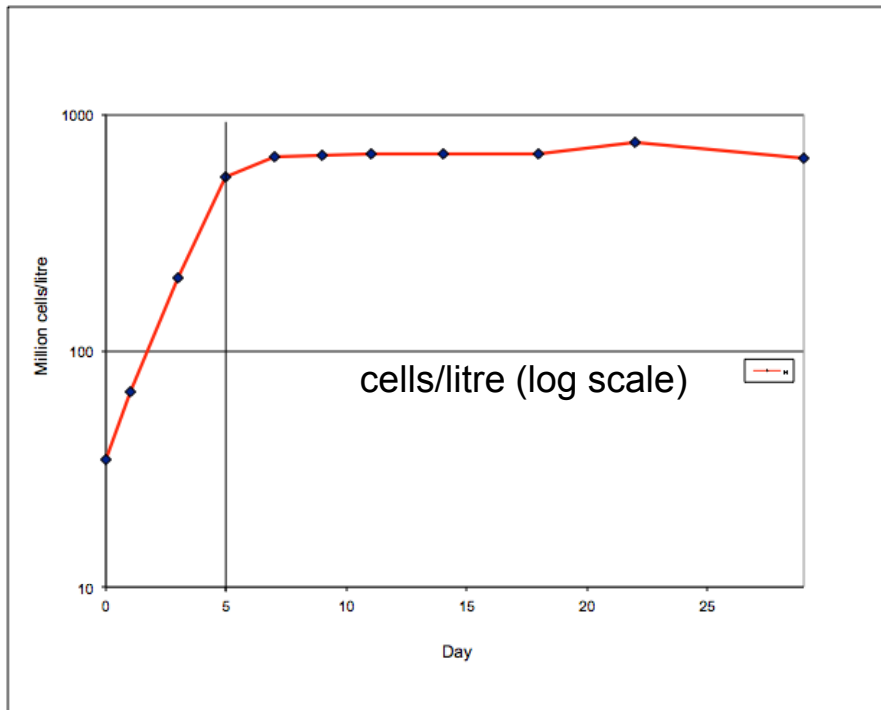
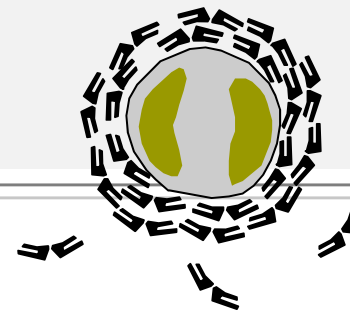
- Hypothesis that varying degree of carbonate saturation has a major effect on coccolithophore ecology and biogeography is not well-supported.
- Extra-cellular calcification may, however, be more vulnerable to changes in saturation state.





# Batch culture

- take sterile nutrient enriched sea-water and add a few thousand cells of *E. huxleyi*



# Cruise-based multi-factorial investigation of the impact of Ocean Acidification on the Pelagic Biosphere

*Jeremy Young, Toby Tyrrell & many others*

## Summary

- 3 major cruises – with objective of developing major datasets to test multiple OA-related hypotheses
- Data synthesis commencing
- Collaboration welcome



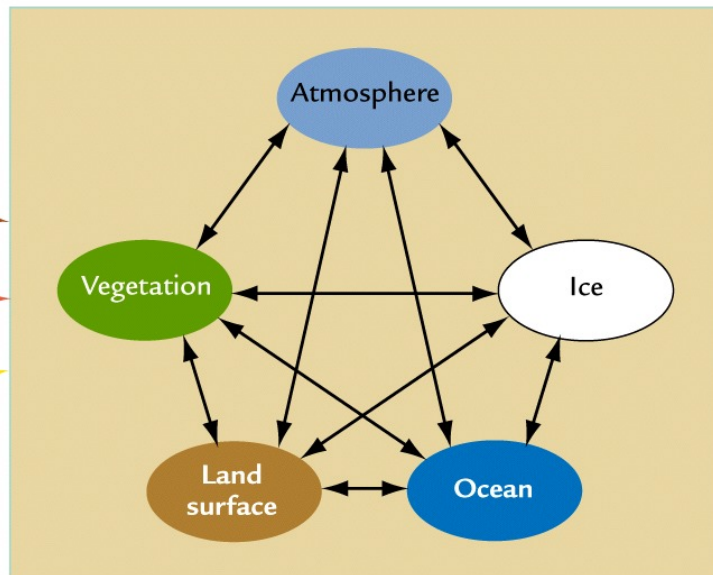
CAUSES  
(external forcing)

Changes in  
plate tectonics

Changes in  
Earth's orbit

Changes in  
Sun's strength

CLIMATE SYSTEM  
(internal interactions)



CLIMATE VARIATIONS  
(internal responses)

Changes  
in  
Atmosphere

Changes  
in  
Ice

Changes  
in  
vegetation

Changes  
in  
Ocean

Changes in  
land  
surface

SIGNALS/PROXIES  
changes in e.g.:

Organism physiology  
(e.g. growth bands)

Faunal compositions

Carbonate chemistry

Sedimentation rates

Earth's climate system - geologist's perspective.

Typical problem is to relate changes in observed geological properties (e.g. changing coccolith assemblage) to external forcings.

